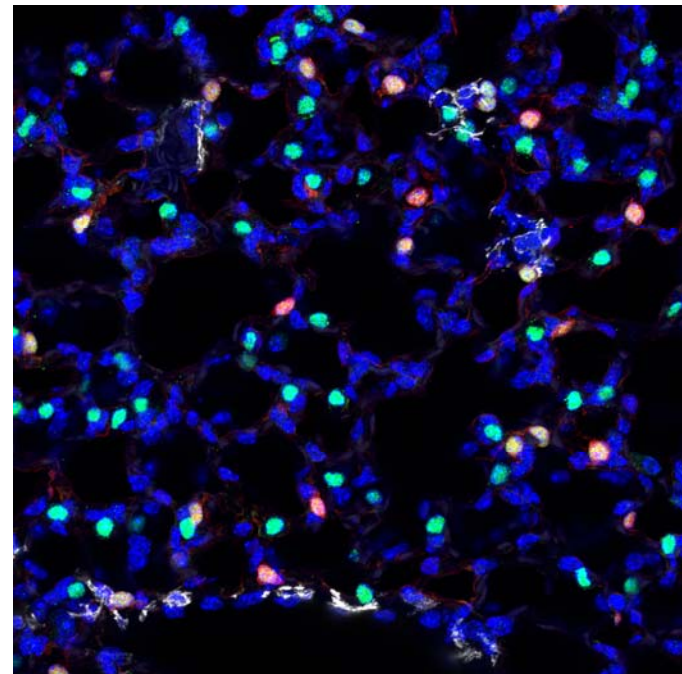
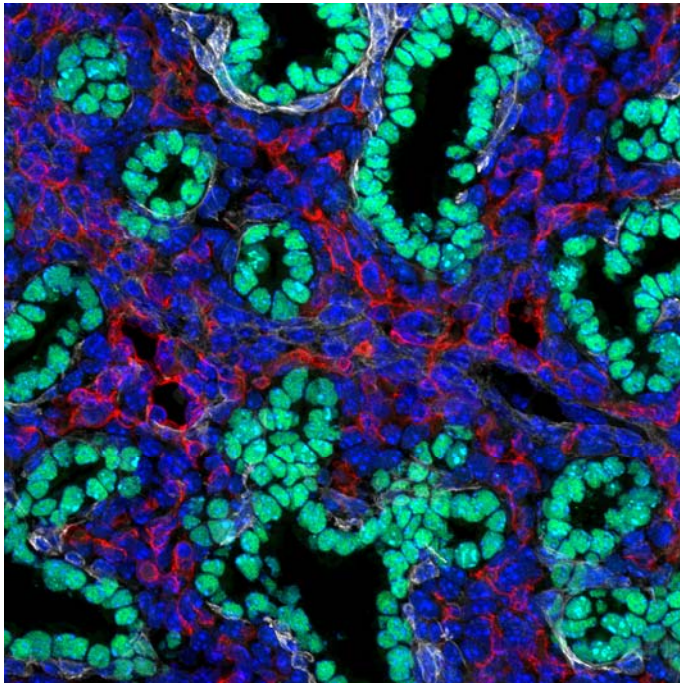
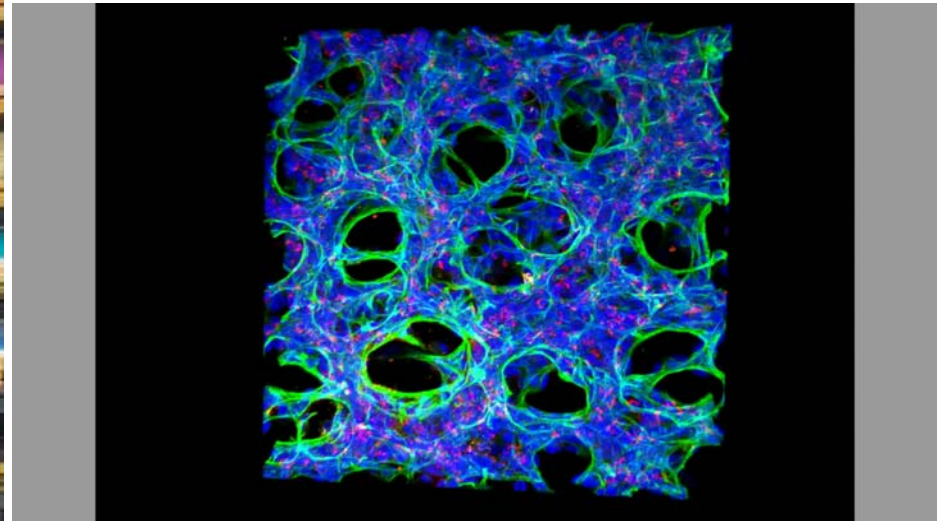


SanDiego 2017:Rare Pediatric Lung Disease
***Single Cell Expression Analysis to Understand
Interstitial Pulmonary Diseases***



Jeff Whitsett: Cincinnati Children's Hospital Medical Center

jeff.whitsett@cchmc.org



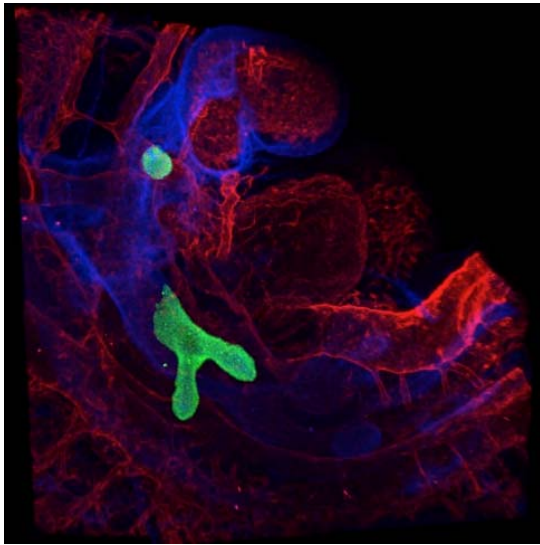


Molecular Tools to Explore Lung Biology

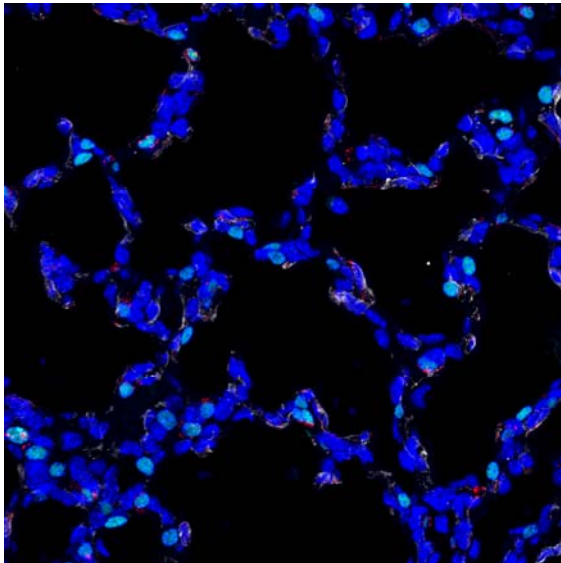


'Maturation' of the Alveolar Epithelium

Lung Buds



PND7



FOXA2, SOX17

E10

E12

Endoderm

Foregut

NKX2-1

Respiratory
epithelium

Proximal Progenitors

SOX2

HOPX/AGER/AQP5

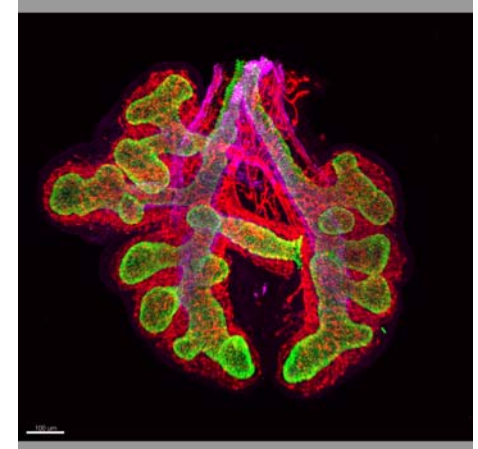
Alveolar
Type I

Alveolar
Type II

Alveoli

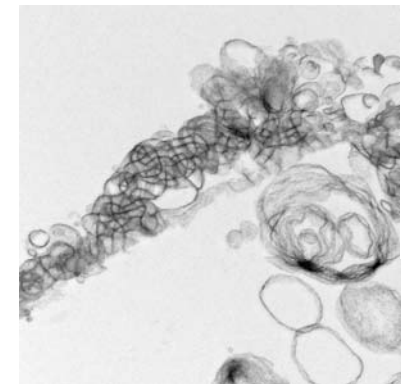


Branching Morphogenesis



Peripheral Progenitors

SOX9, NKX2-1

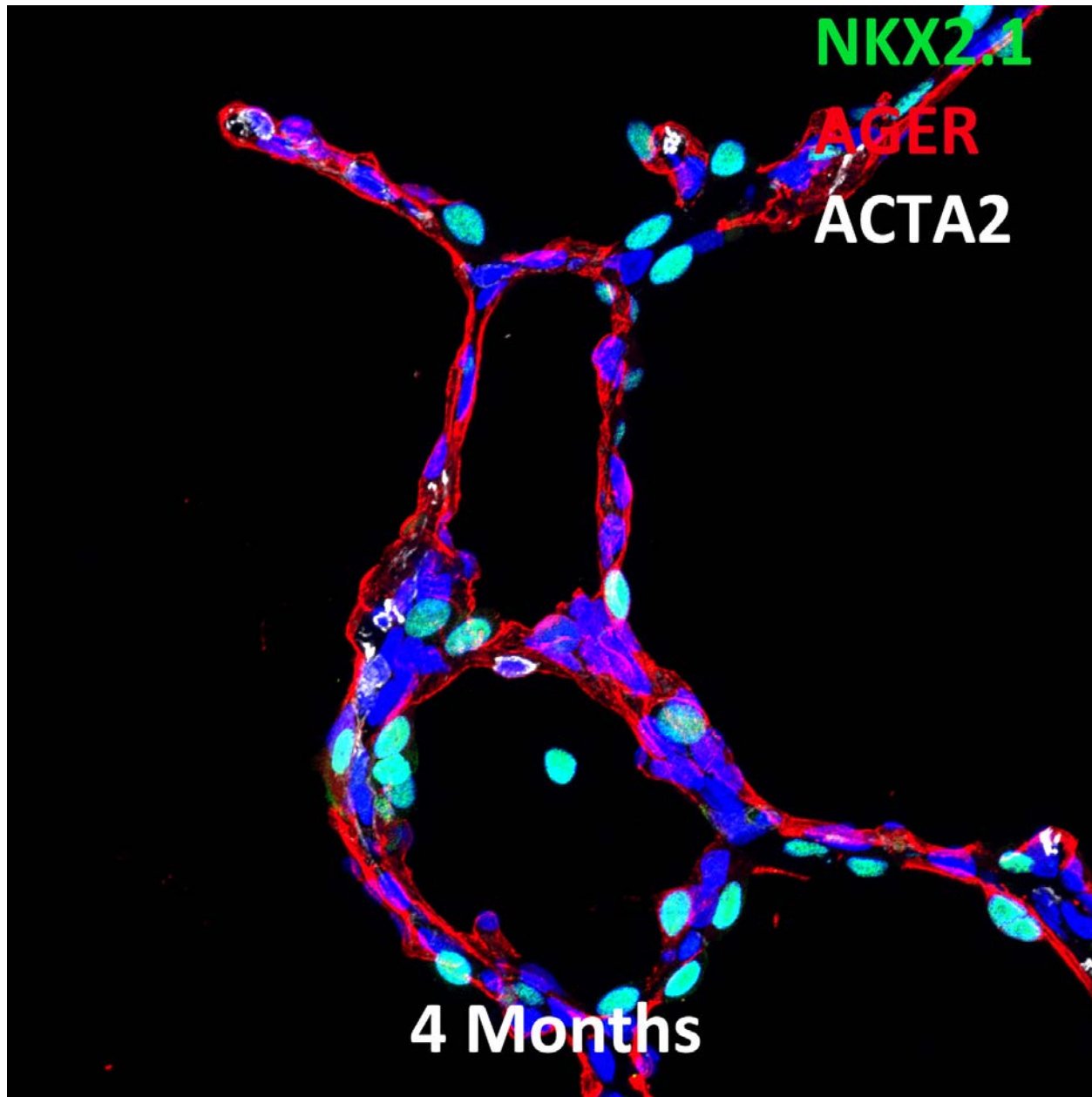


Alveolar Surfactant

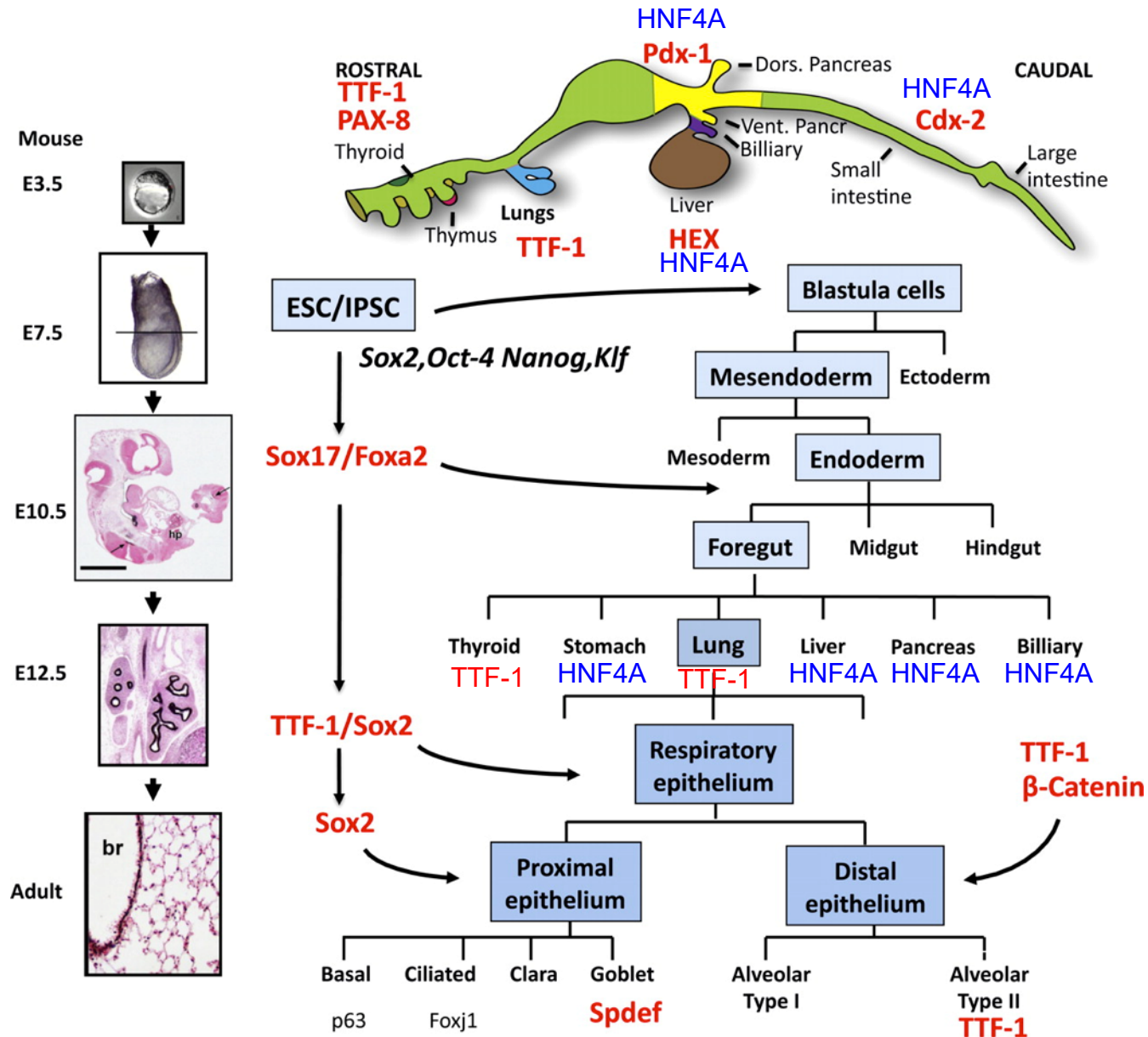
SFTPA/B/C/D

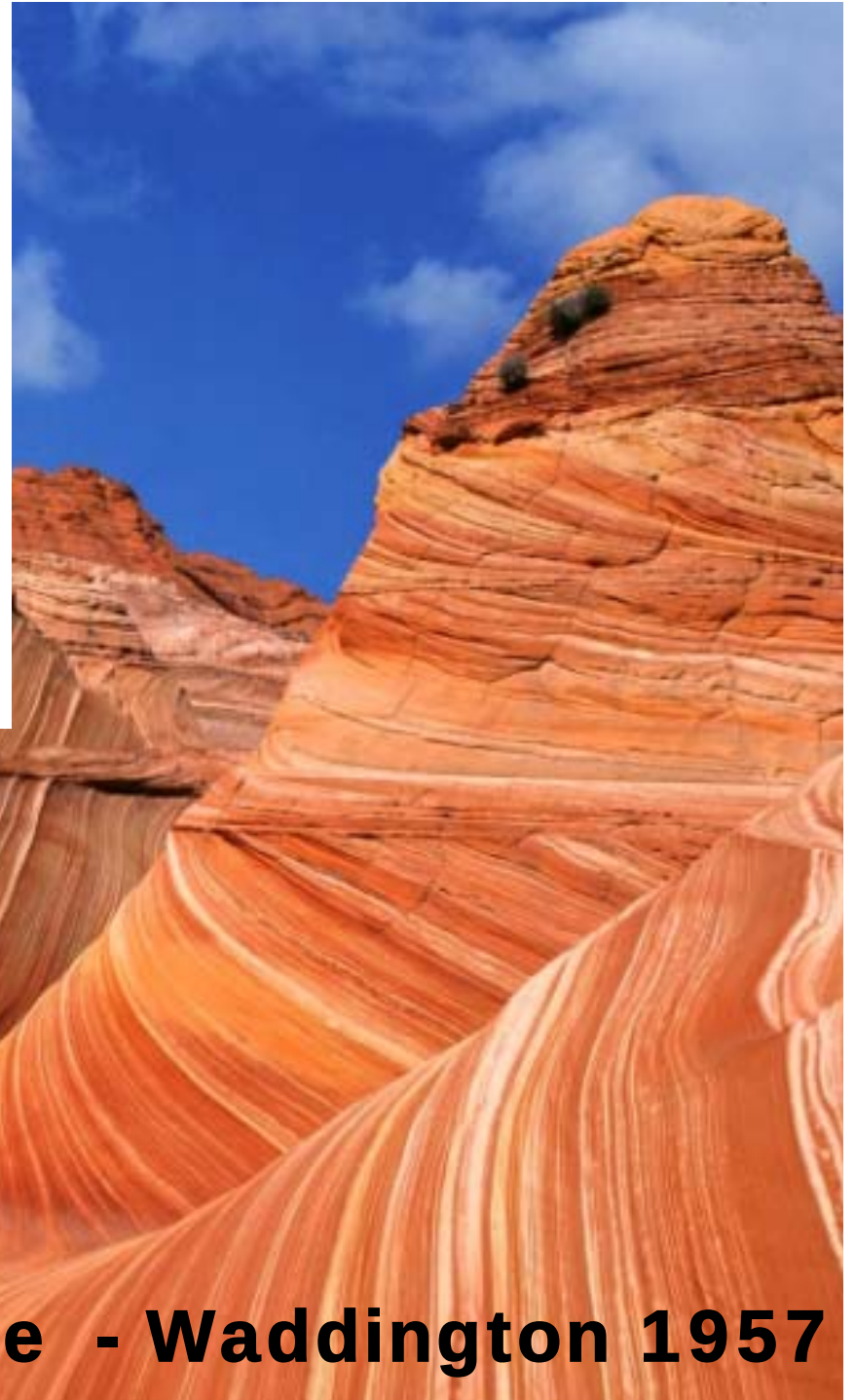
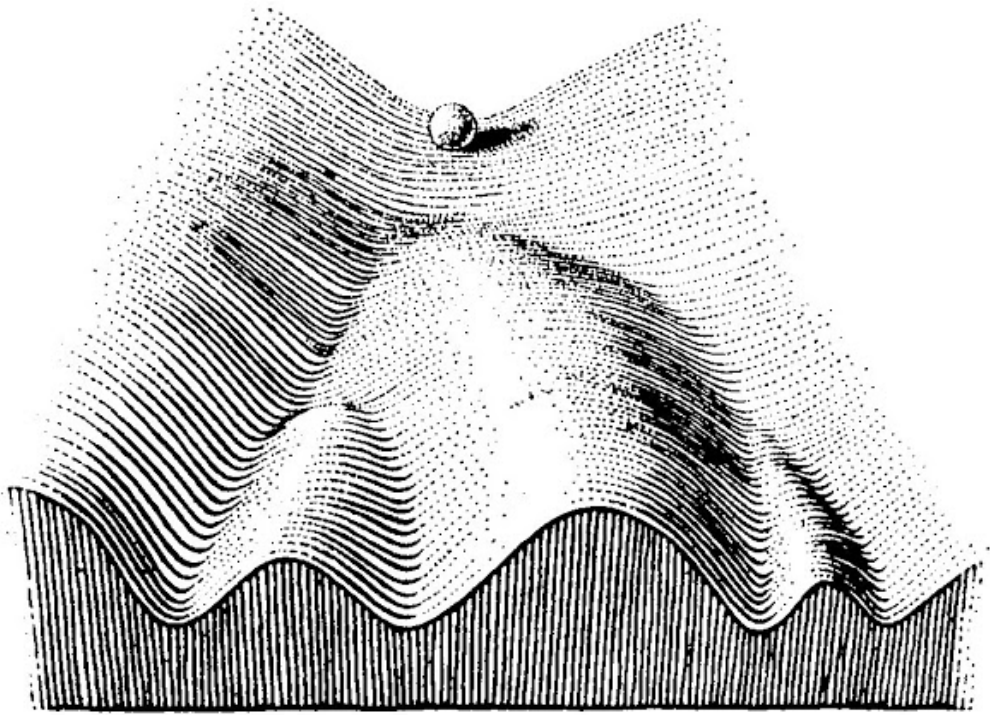
ABCA3, GPR116, NAPSA

*Alveolar AT1 [**AGER**], AT2 [**NKX2-1**] and Myofibroblasts [ACTA2] in Human Lung*



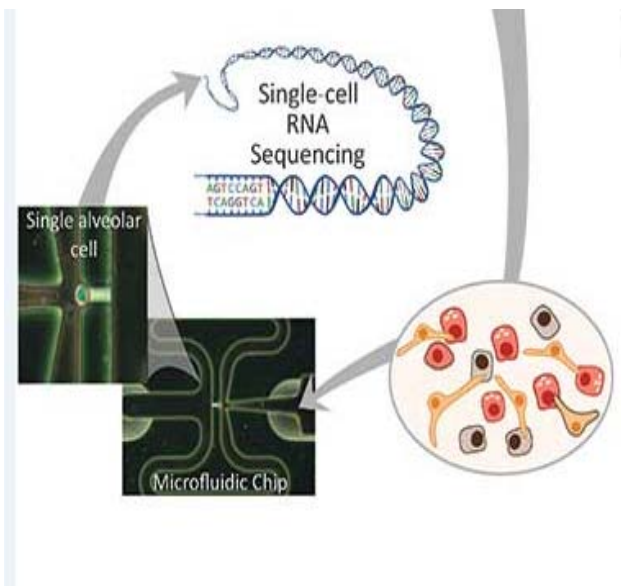
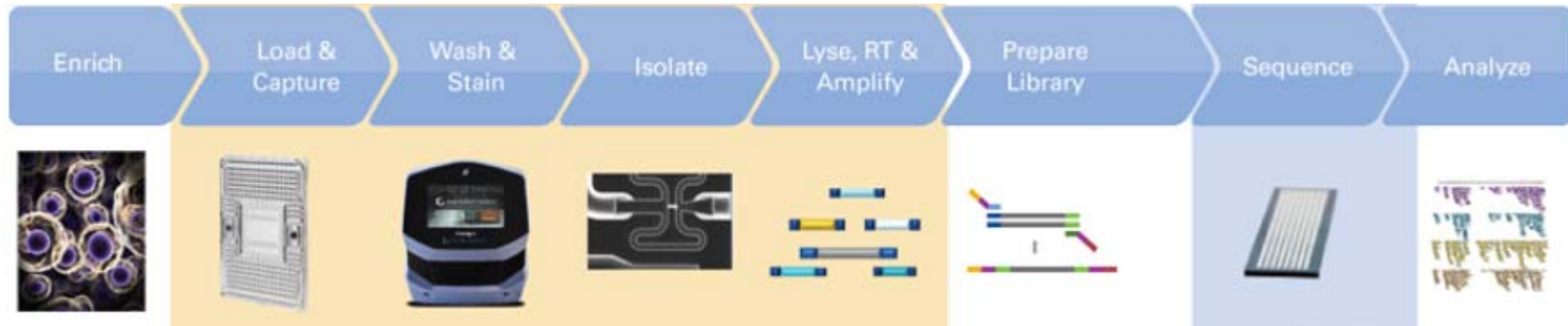
TTF-1/NKX2-1 keeps lung integrity by activating lung specific genes and suppressing liver specific genes





The Epigenetic Landscape - Waddington 1957

Single Cell RNA-Seq



isolate single cells from protease-dispersed lungs

↓

separate them using the Fluidigm C1™ Single-Cell Auto Prep System

↓

convert RNA into a sequencing library using Fluidigm PN100-5989

↓

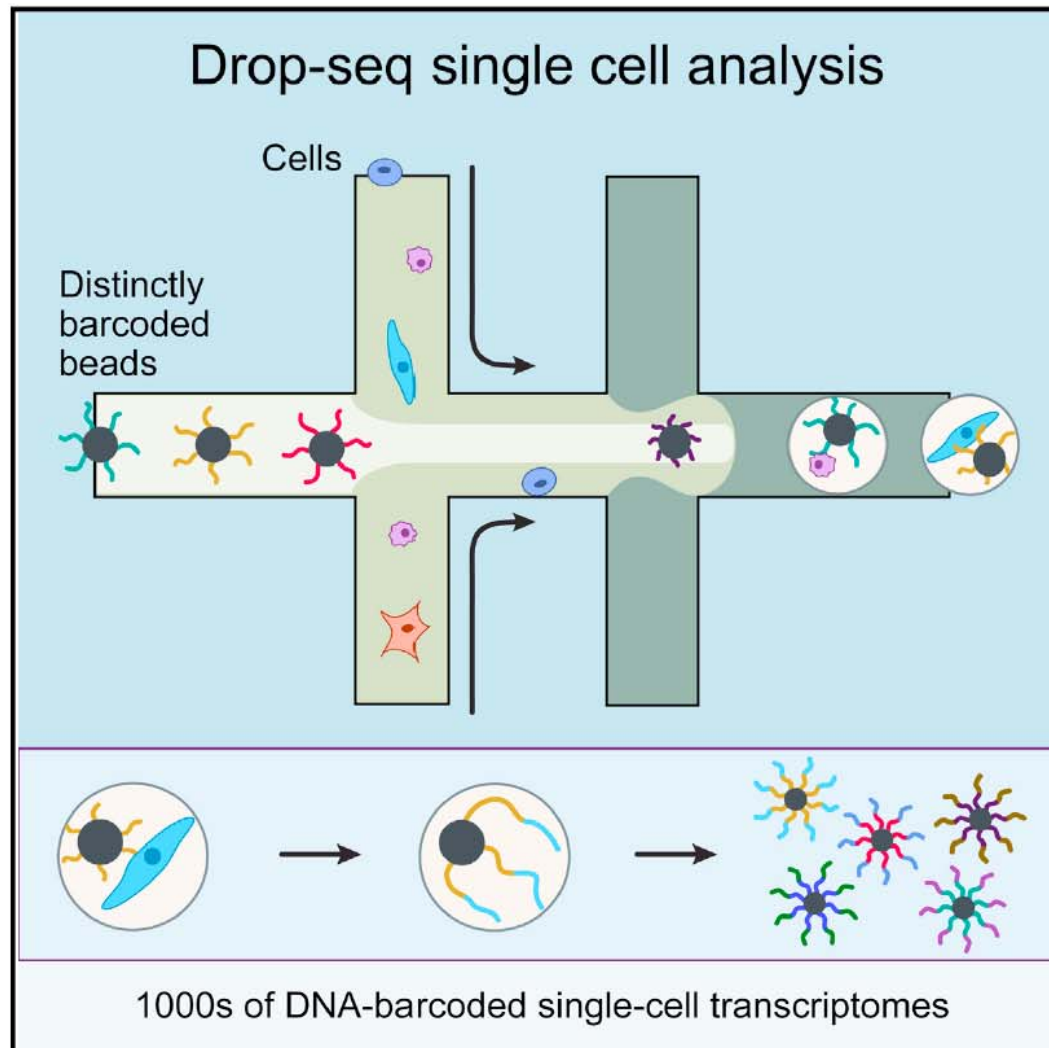
RNA-Seq on 96 individual lung cells per run on Illumina 2500 Hi-Seq

↓

RNA-seq data QC, alignment and analysis

Highly Parallel Genome-wide Expression Profiling of Individual Cells Using Nanoliter Droplets

Graphical Abstract



Authors

Evan Z. Macosko, Anindita Basu, ..., Aviv Regev, Steven A. McCarroll

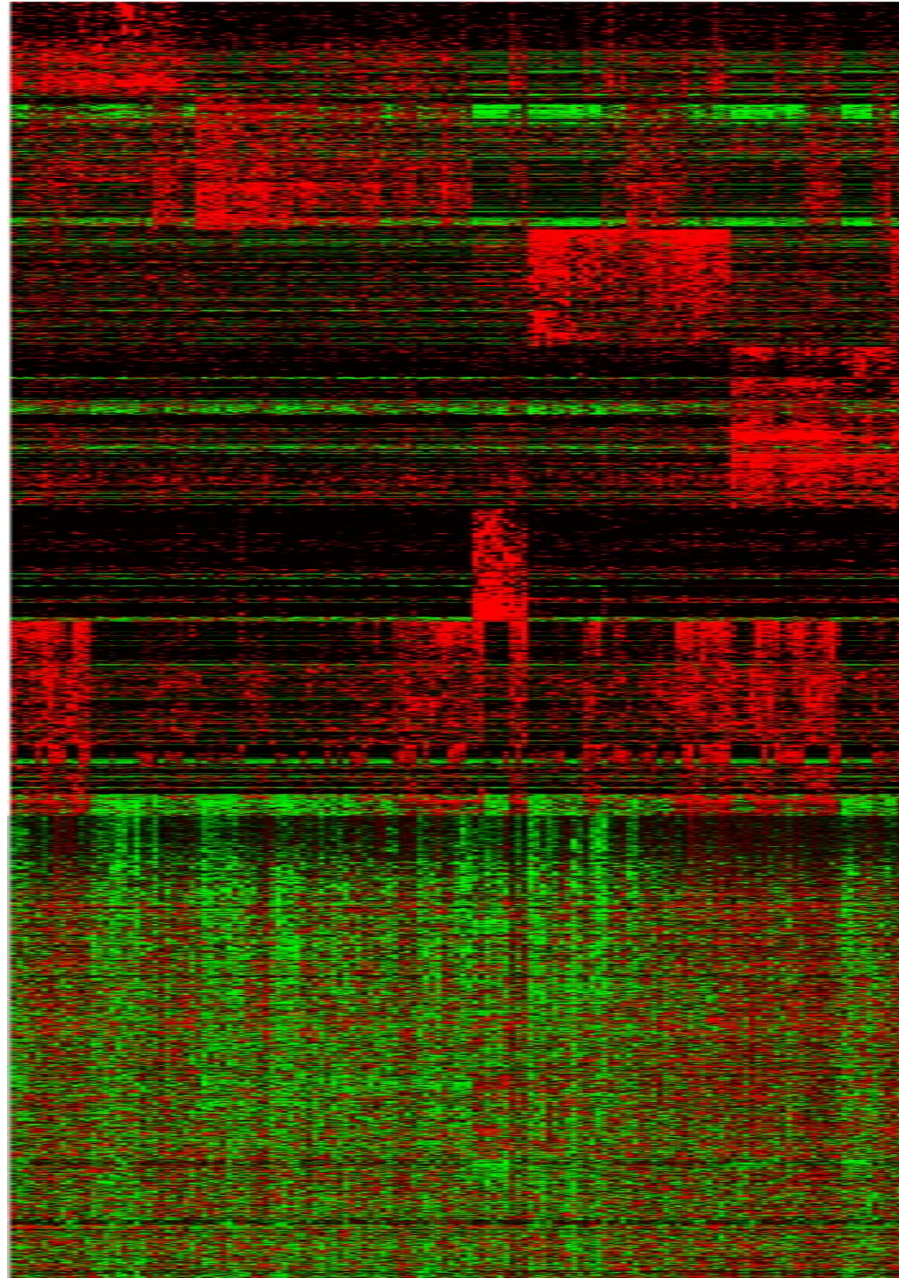
Correspondence

emacosko@genetics.med.harvard.edu (E.Z.M.),
mccarroll@genetics.med.harvard.edu (S.A.M.)

In Brief

Capturing single cells along with sets of uniquely barcoded primer beads together in tiny droplets enables large-scale, highly parallel single-cell transcriptomics. Applying this analysis to cells in mouse retinal tissue revealed transcriptionally distinct cell populations along with molecular markers of each type.

4232 Genes
(compendium of
clusters



C1 Myo- fibroblast

C2 Fibroblast

C3 Endothelial

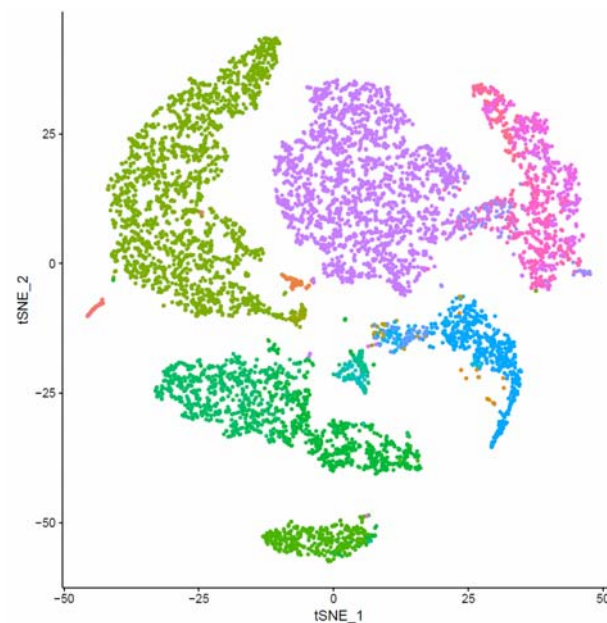
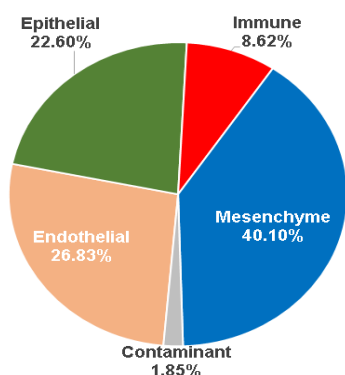
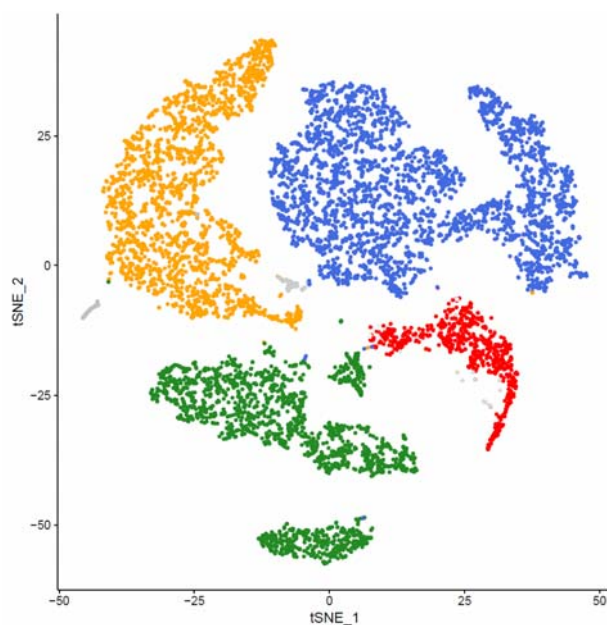
C4 Epithelial

C5 Myeloid

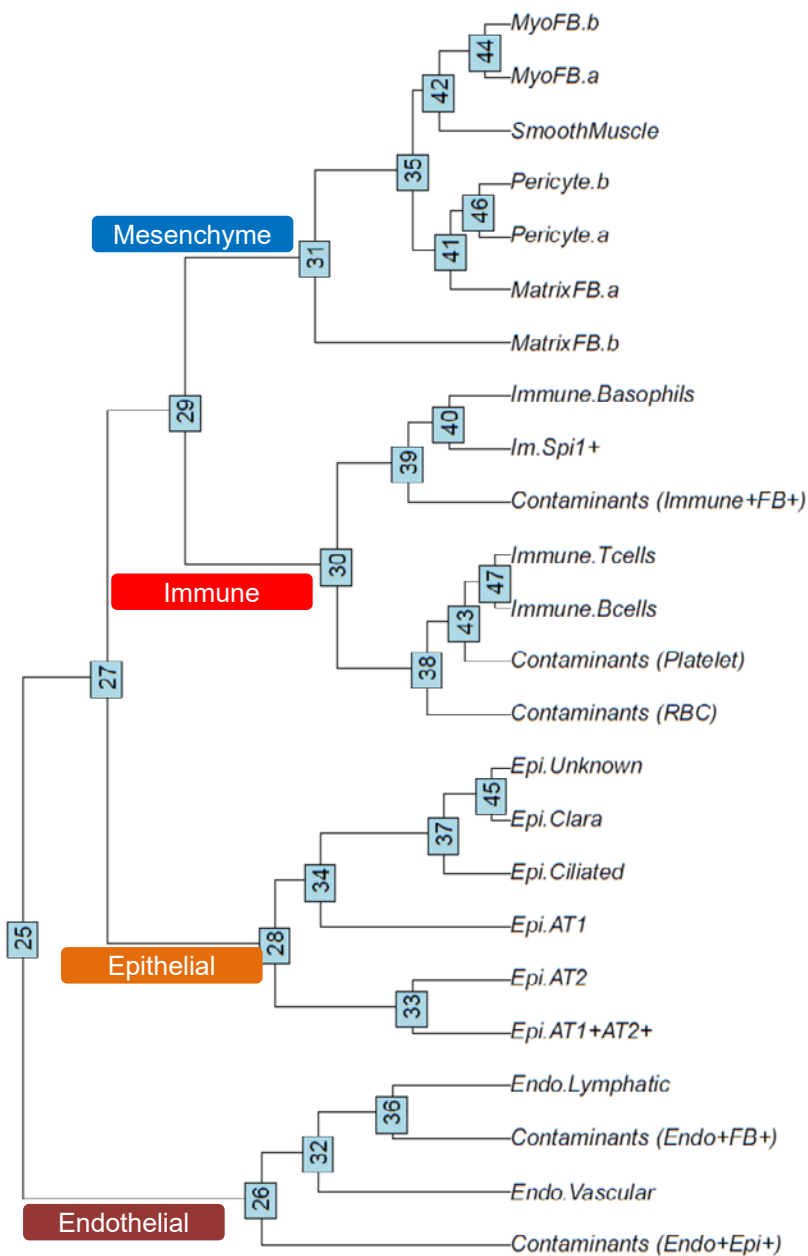
C6 Cell Cycle

C7 Growth / Housekeeping

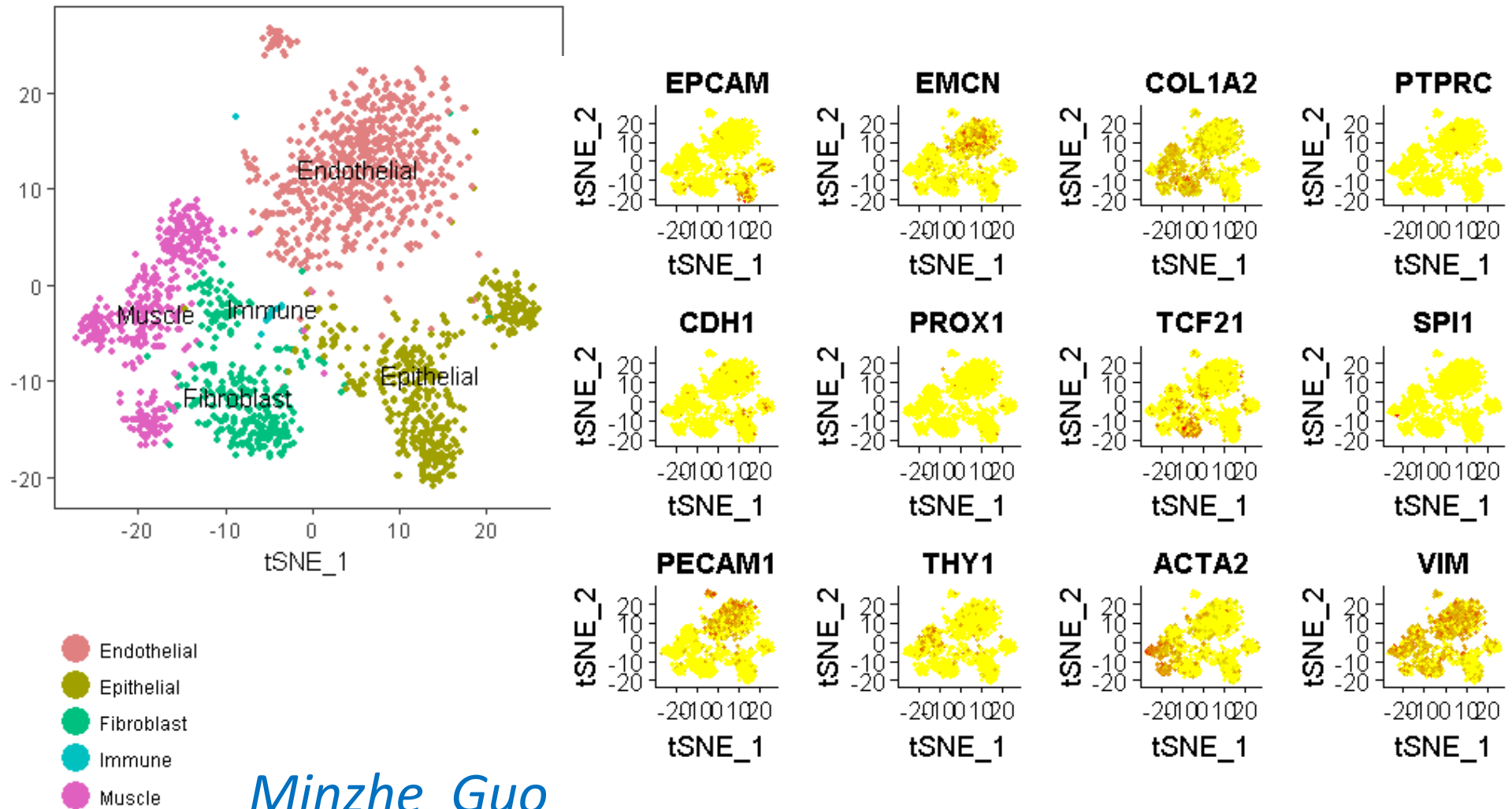
Mouse P1 Cell Types Identification



19 Subtypes



Human Infant-DAY1 Drop-seq: Major Cell Types and Marker Expression (HTC)

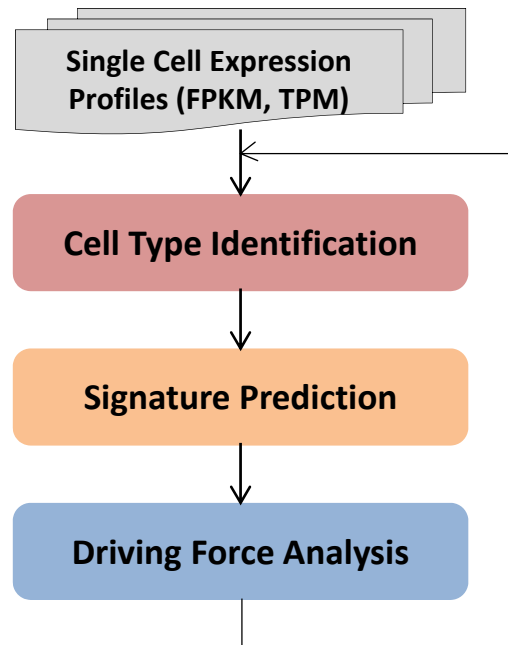


Minzhe Guo

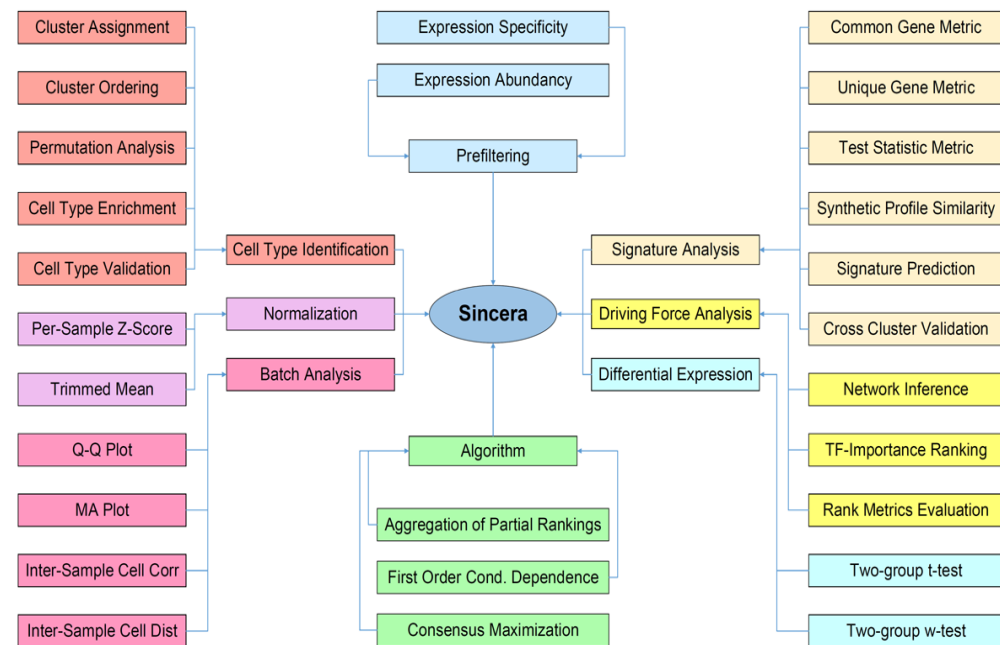
CCHMC/U of Rochester

Sincera: a computational pipeline for SINgle CELL Rna-seq profiling Analysis

Main Analysis Components



Program Structure of Sincera



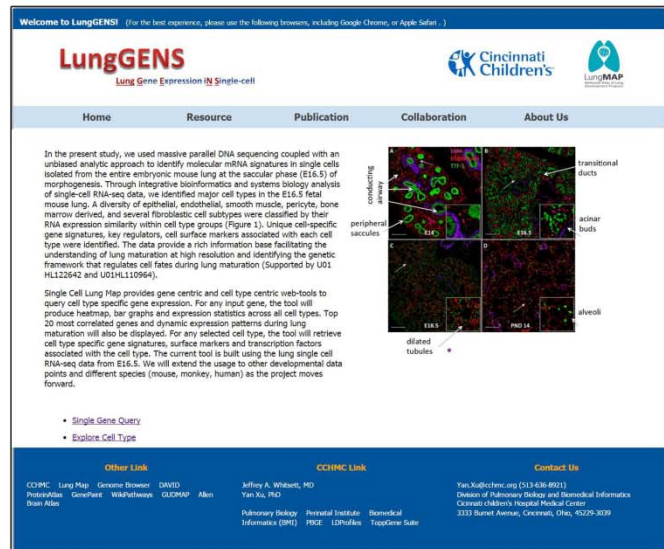
<https://research.cchmc.org/pbge/sincera.html>

Screenshot of LungGENS

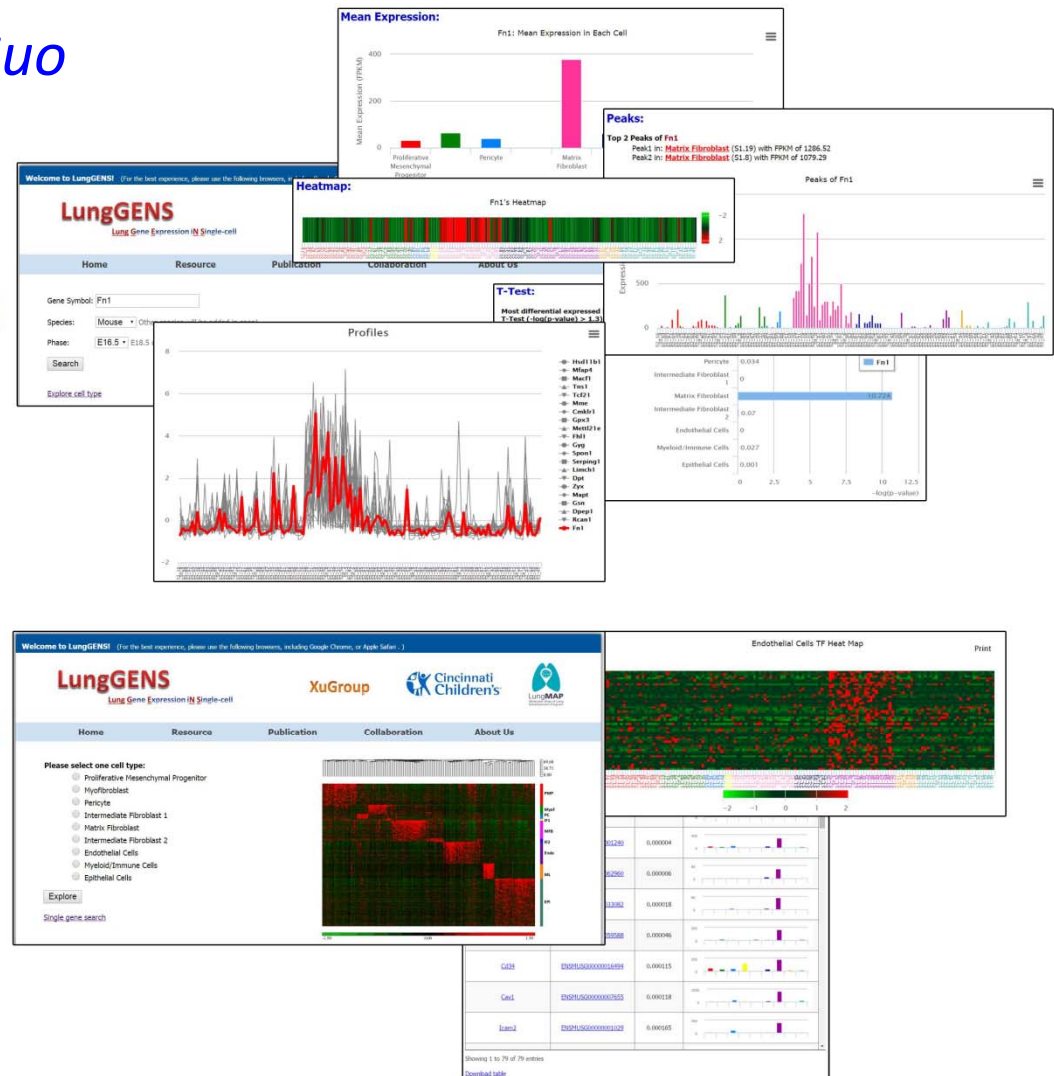
(<https://research.cchmc.org/pbge/lunggens/default.html>)

Yan Xu, Yina Du, Mizhe Guo

Query by Gene



Query by Cell Type



CCHMC

LGEA Web Portal

Lung Gene Expression Analysis Web Portal



LGEA Web Portal

LungGENS

Lung Image

Collaboration

News

LGEA Tools

We previously developed "LungGENS", a web site for mapping single cell gene expression in the developing lung. Initial phases of the webtool were built based on single cell RNAseq data from normal fetal mouse lung at E16.5. Web statistics indicated that "LungGENS" is well received and visited by research investigators worldwide. With the continued support from the "LungMAP" consortium, transcriptome data derived from different technique platforms, species and lung development stages are quickly expanding. We now extend the scope and contents of the LungGENS database to accommodate heterogeneous data structures and types. We present "LGEA web portal" (Lung Gene Expression Analysis web portal), an extended version of LungGENS web portal for analyses of lung gene expression patterns and mapping cell types in the developmental lung from single cells, sorted cell populations and whole tissue developmental studies.

LGEA implements a number of new features and analytical methods. It allows the analysis of 1) single cell transcriptomes using **LungGENS**; 2) sorted lung cell populations using **LungSortedCells**; 3) Lung Developmental Time Course analysis using **LungDTC** and 4) Lung Disease analysis **LungDisease**.

News and updates:

- **Introduction of LungMAP** -- Dr. Jeffrey Whitsett
- **5-8-2017**: LGEA added in [mouse P7 scRNA-seq dataset](#) and [Human IPSC-derived NKX2-1 lung scRNA-seq dataset](#) from Kotton Lab.
- **4-22-2017**: Dr. Whitsett and Dr. Xu's new published chapter: [Transcriptional Networks-Control of Lung Maturation](#).
- **4-22-2017**: Check out our 3 new sections: "LungEpigenetics", "LungImage" and "LGEA-ToolBox".
- **4-10-2017**: LGEA web portal has just been published by [Thorax](#). ([PubMed](#))
- **6-9-2016**: Check out this new book "Fetal and Neonatal Lung Development: Clinical Correlates and Technologies for the Future (Lung Growth, Development, and Disease)" (1st Edition by [Alan Jobe](#) (Editor), [Jeffrey Whitsett](#) (Editor), [Steven Altmann](#) (Editor)). ([Amazon](#))

<<More>>

**If you are experiencing problems displaying LGEA pages, please refresh the page or clear the browser cache.

LungGENS

LungGENS - species

Mouse (scRNA-seq, drop-seq)
(E16.5, E18.5, P1, P7, P28) [new](#)

Monkey
(Day 90, TGA)

LungGENS - query

Query by single gene
Query by cell type
Query by gene list

LungSortedCells

Human

Human Sorted (Immune, Endothelial,
Mesenchymal and Epithelial Cells)

Mouse

Mouse AT2
(P19, P19.28)

Mouse Pdgfra+ Fibroblast
(E18.5, E18.5/P19, P19.28)

Mouse Sorted (Immune, Endothelial and
Epithelial Cells) [new](#)

LungSortedCells-query

Query by gene list

LungDTC

MicroArray

LDProfiles
MGI: developmental time course

RNA-seq

Mouse Lung
(E16.5, E18.5, P1, P3, P7, P14 and P28)

Rhesus Macaque Lung
(GA105, GA130, GA150)

LungDisease

scRNA-seq Analysis

Human IPF
(S40 normal and disease cells)

Human IPSC-derived NKX2-1+ lung
(Kotton Lab normal and CF)

Sorted RNA-seq

Human IPF
Human AT2 cells

LungEpigenetics

ENCODE

Mouse histone PTM chip-seq
data [new](#)

LungMAP research center

TBA

LungImage

Confocal Imaging

Mouse C57BL6 lung
(E18.5, E18.5, P1, P3, P7, P10...) [new](#)

Human Lung
(Fetal and Adult) [new](#)

H&E Staining

Human Lung
(Fetal and Adult) [new](#)

Query by cell type

TBA

LGEA-ToolBox

Database query tool

Signatures comparison
Gene at glance
LDProfiles

Research tools

SINCERA

SLICE

Other resource

Tool list

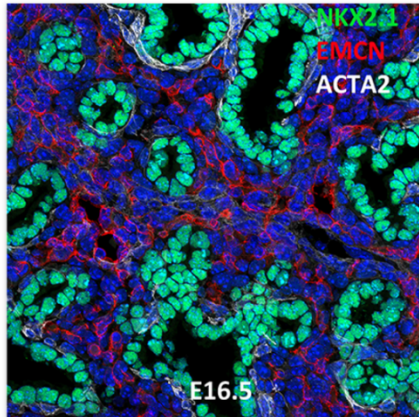
Current version and under development

<https://research.cchmc.org/pbge/lunggens/mainportal.html>

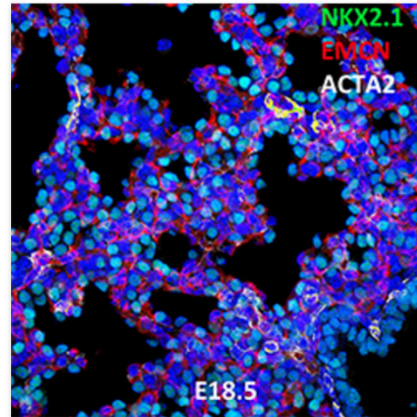
<https://www.lungmap.net/>

“LGEA “and “Breath”:Atlas of Mouse Lung Cell Types During Development

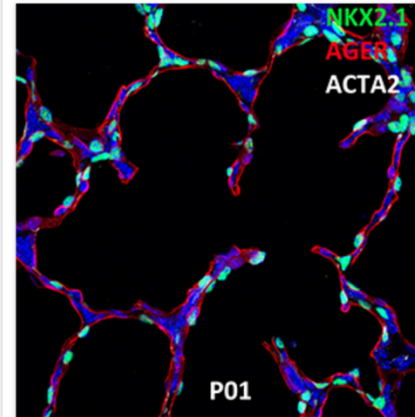
C57BL/6



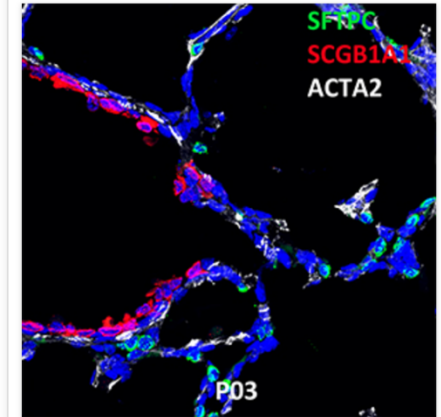
E16.5 C57BL/6 Confocal Imaging



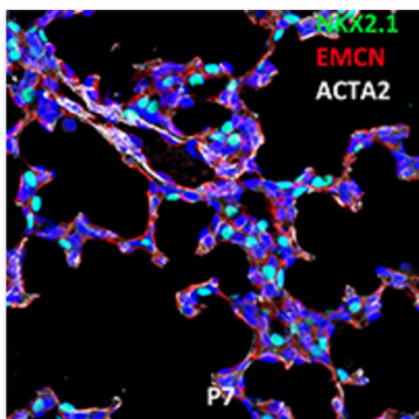
E18.5 C57BL/6 Confocal Imaging



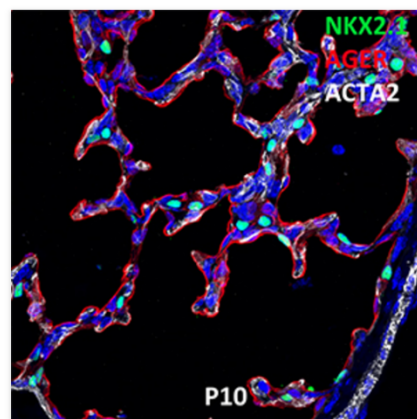
Postnatal Day 1 C57BL/6 Confocal Imaging



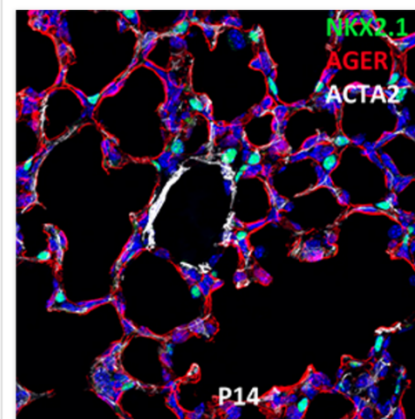
Post Natal Day 3 C57BL/6 Confocal Imaging



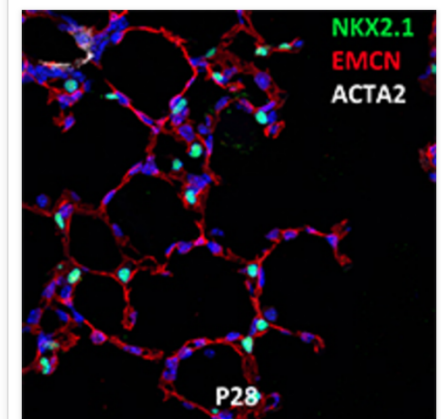
Postnatal Day 7 C57BL/6 Confocal Imaging



Postnatal Day 10 Confocal Imaging

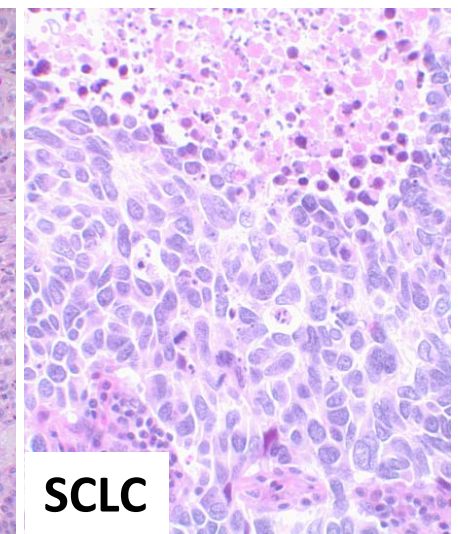
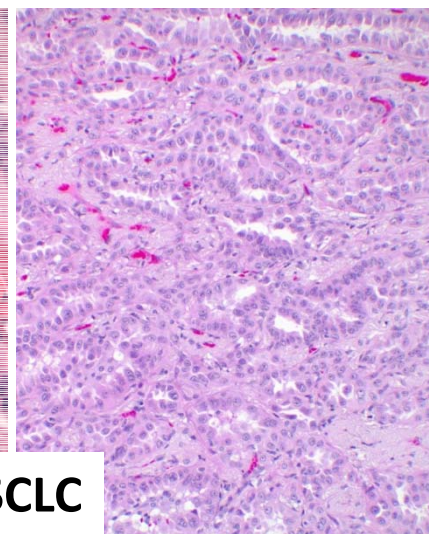
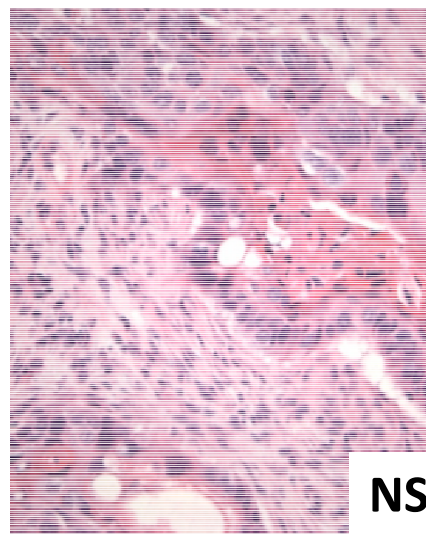
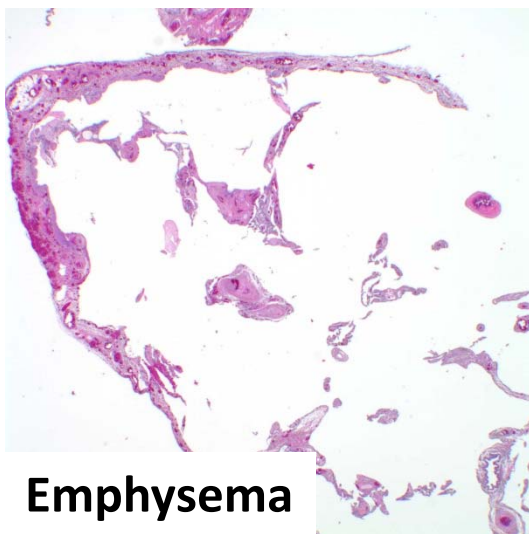
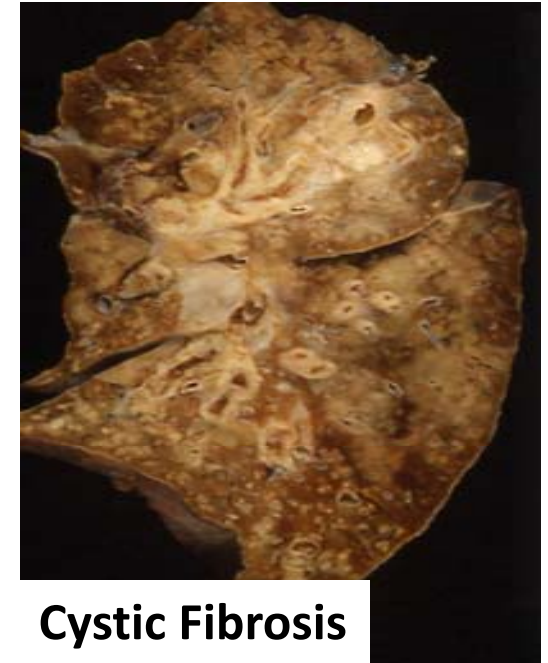
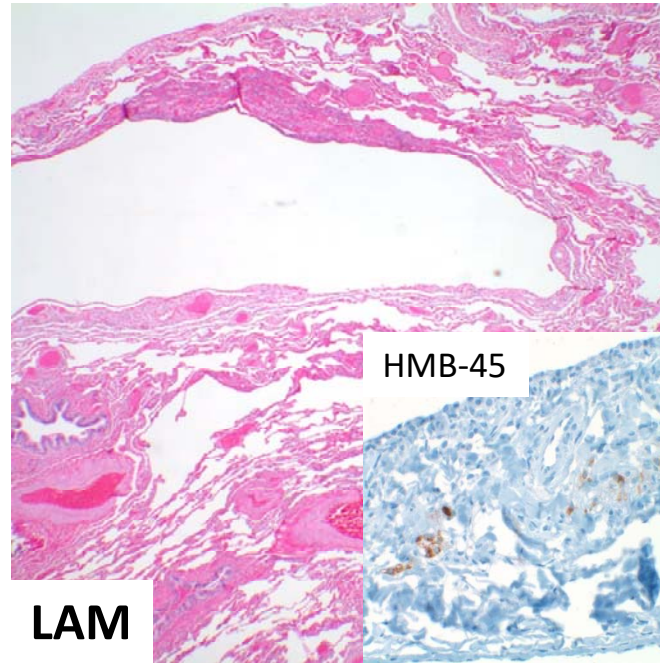
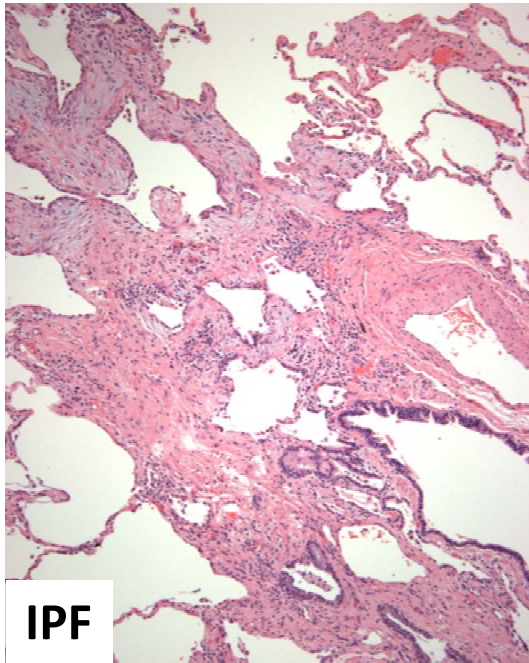


Postnatal Day 14 C57BL/6 Confocal Imaging



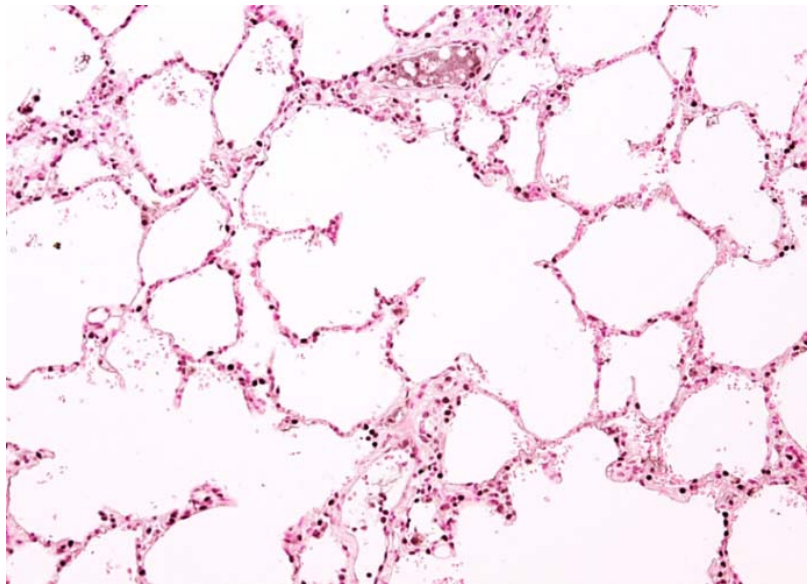
Postnatal Day 28 C57BL/6 Confocal Imaging

Challenge of Chronic Pulmonary Diseases

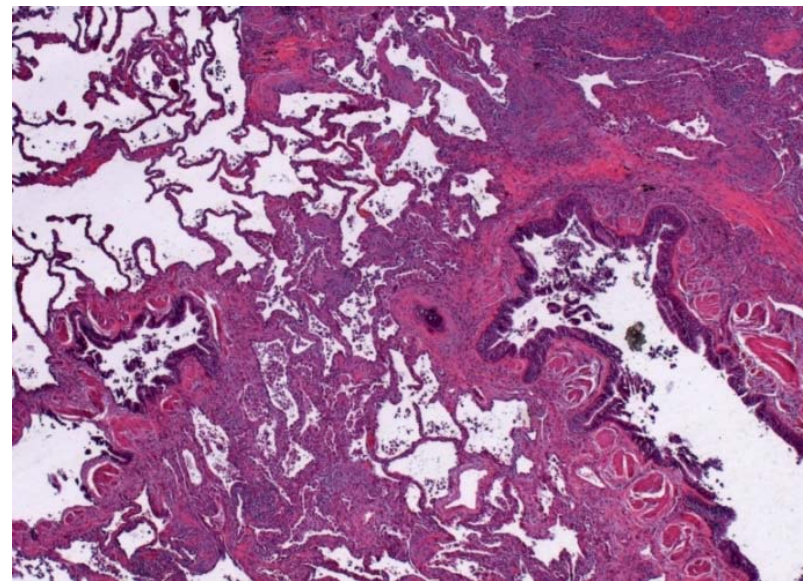


Idiopathic Pulmonary Fibrosis

- Normal

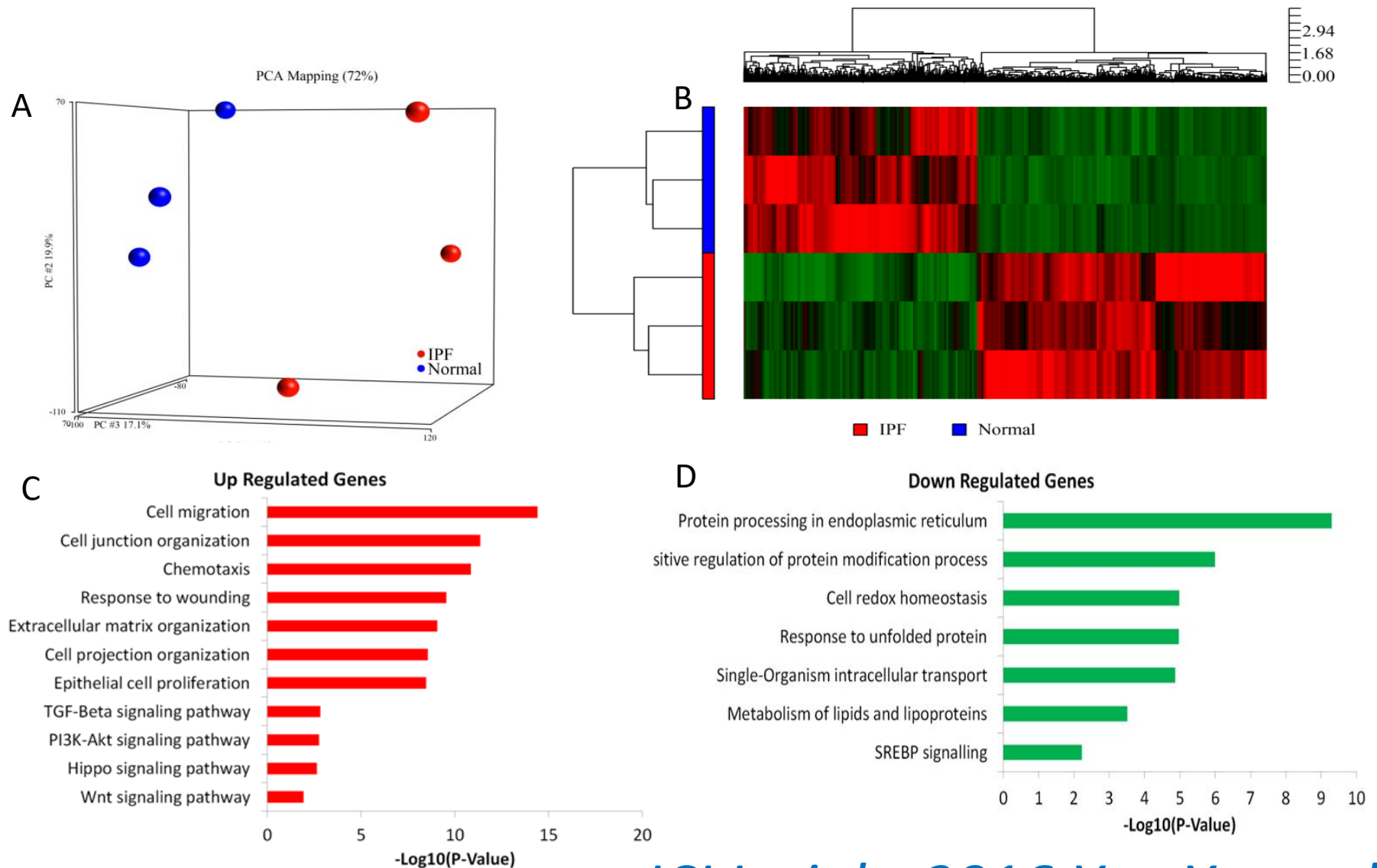


IPF



Barry Stripp-Cedar Sinai

Gene Expression in Human IPF :Sorted Epithelial Cells

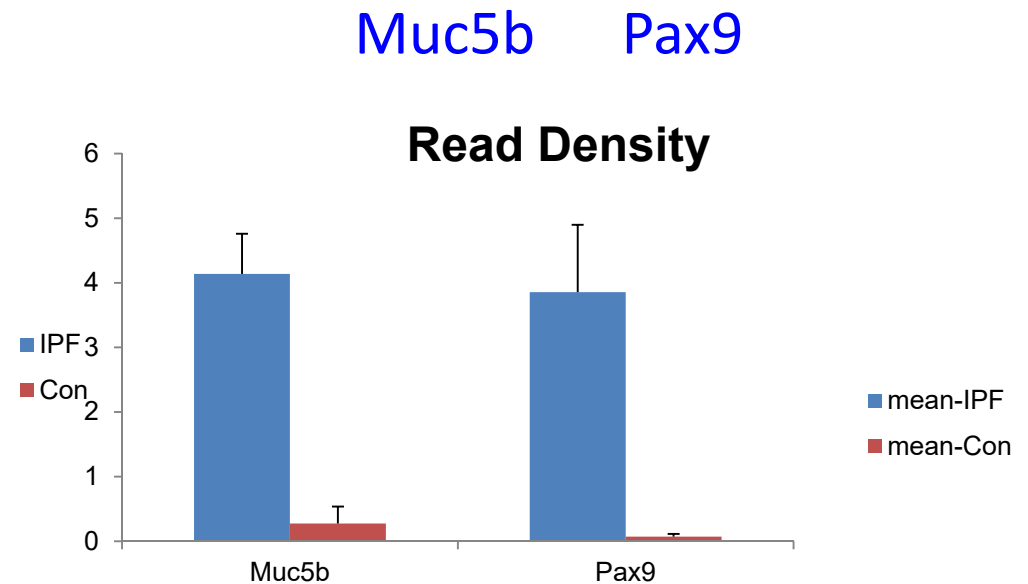
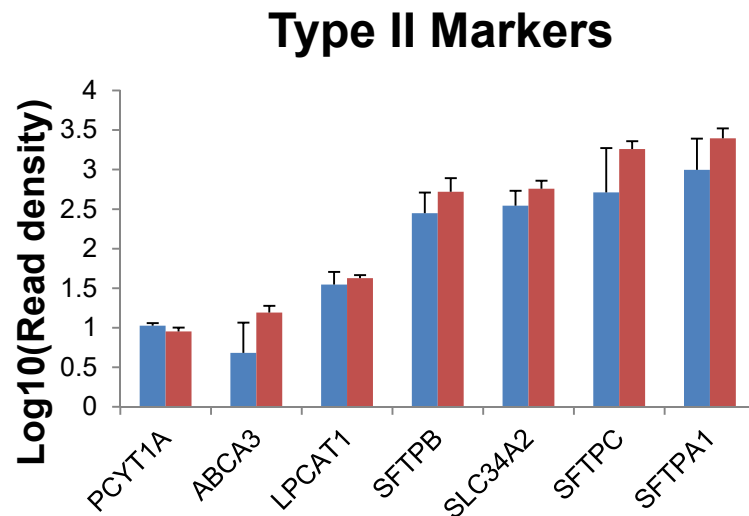


JCI Insight 2016;Yan Xu et al

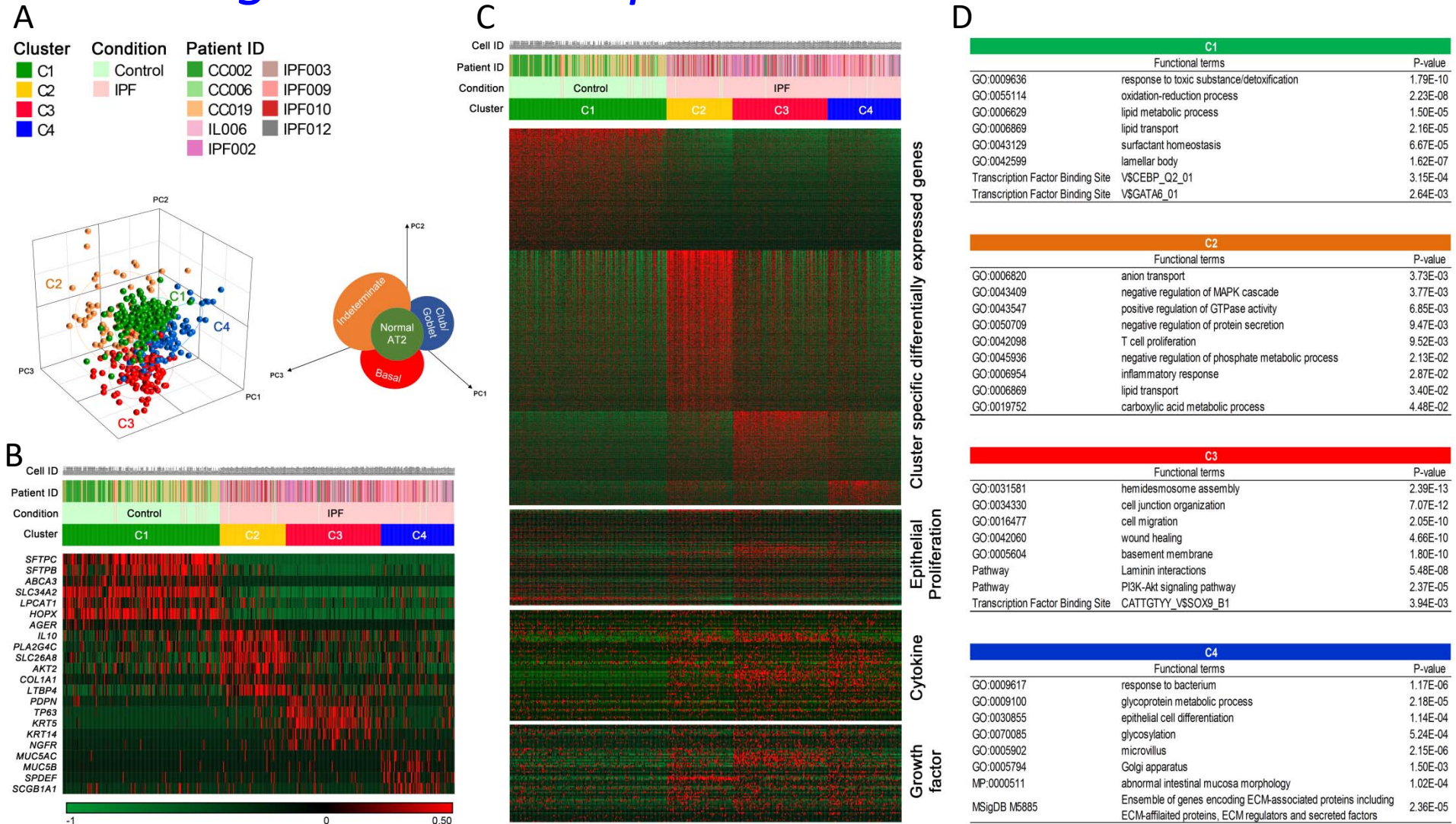
Expression of Proximal Epithelial Markers in IPF Type 2 Cells

Loss of Normal Epithelial Cell Identity

SFTPA1, SFTPB, SFTPC, PCYT1A,
ABCA3, SLC34A1, LPCAT1

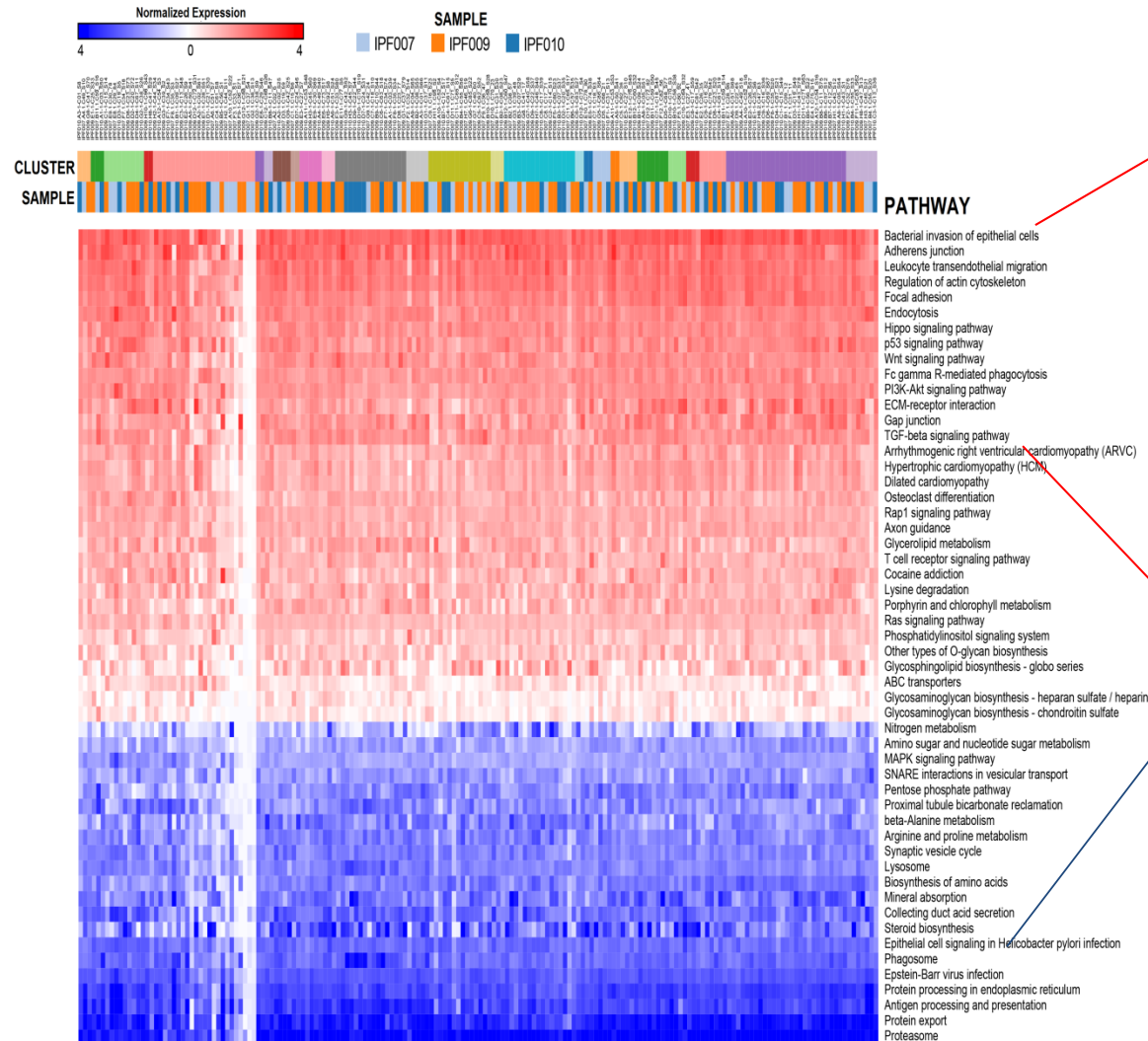


Single Cell RNA Expression Patterns in IPF



JCI Insight 2016/Yan Xu et al

Enrichment of KEGG Pathway in IPF SC

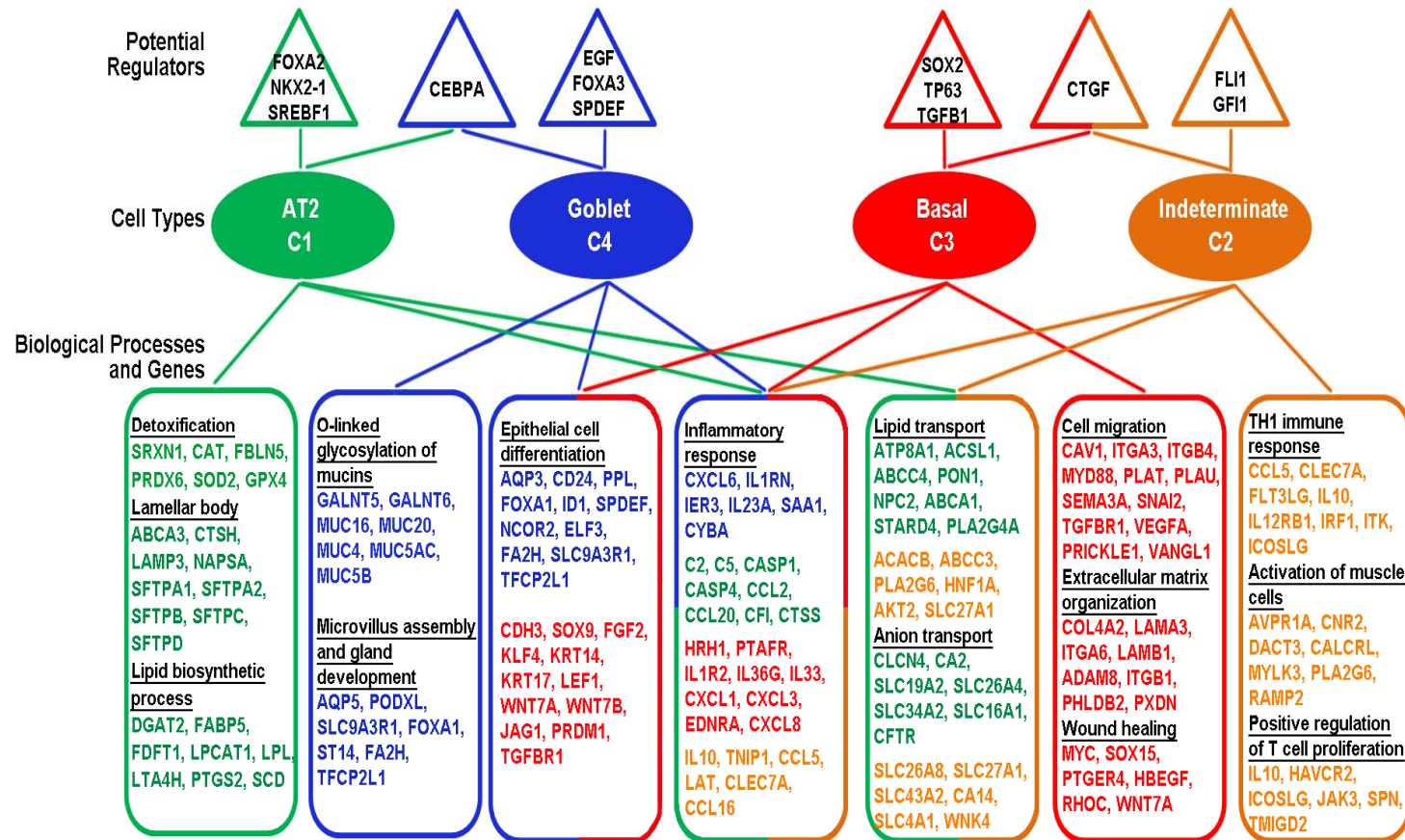


- Bacterial invasion of epithelial cells
- Adherens junction
- Leukocyte transendothelial migration
- Regulation of actin cytoskeleton
- Focal adhesion
- Endocytosis
- Hippo signaling pathway
- p53 signaling pathway
- Wnt signaling pathway
- Fc gamma R-mediated phagocytosis
- PI3K-Akt signaling pathway
- ECM-receptor interaction
- Gap junction
- TGF-beta signaling pathway

- Proteasome
- Protein export
- Antigen processing and presentation
- Protein processing in endoplasmic reticulum
- Epstein-Barr virus infection
- Phagosome

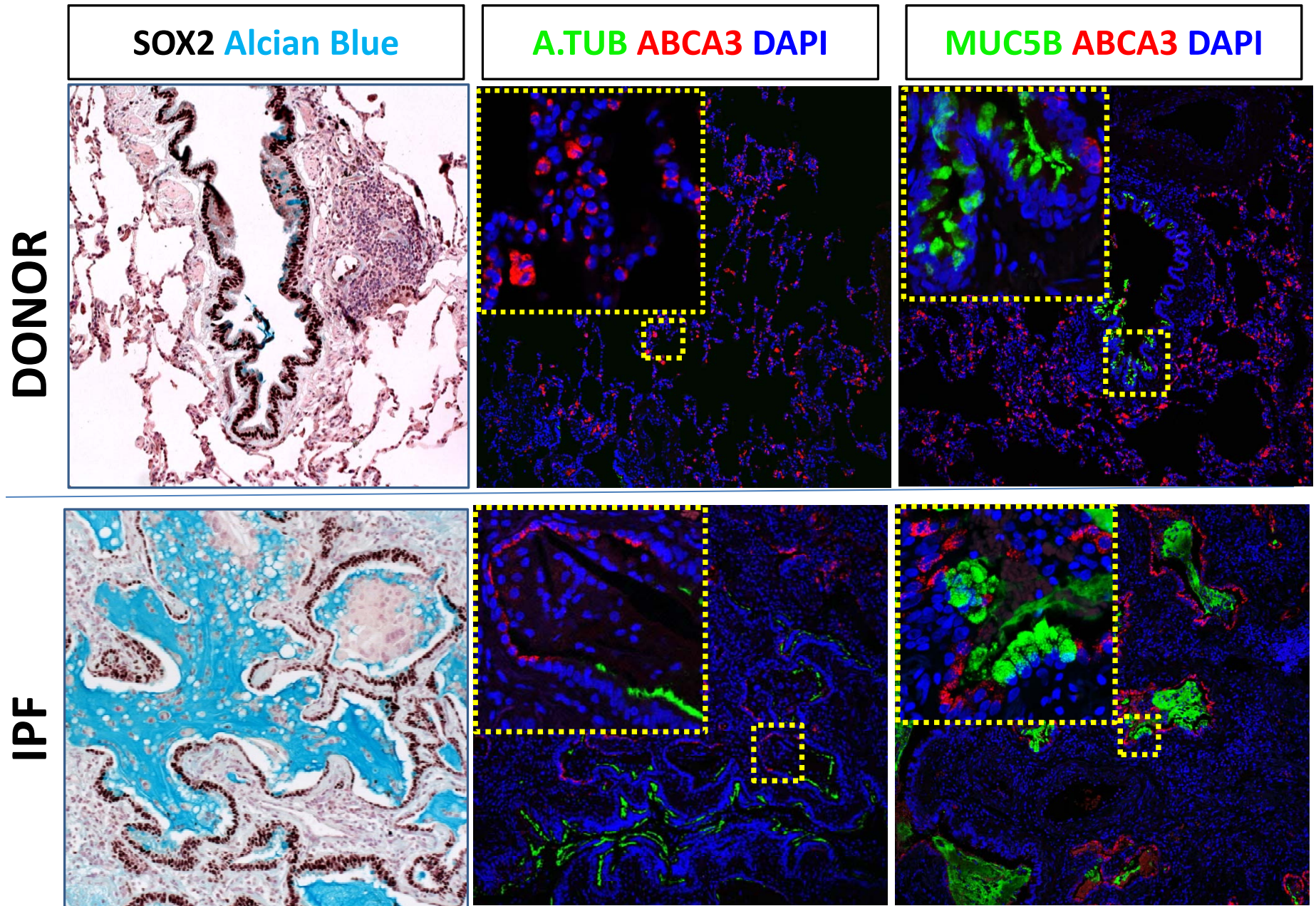
Normal AT2 Cells

IPF Epithelial Cells

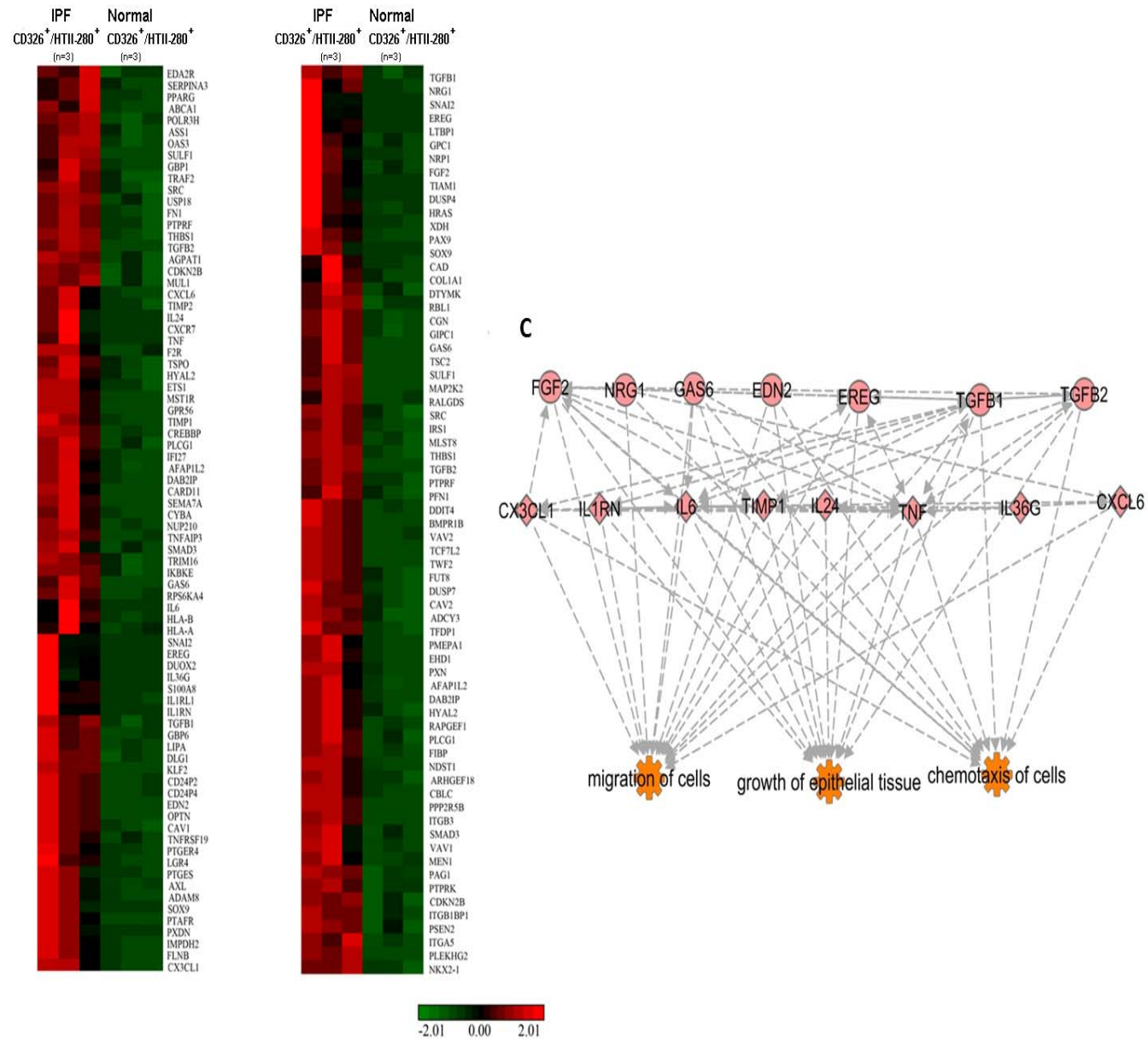


Loss of Alveolar Cell Identity in IPF

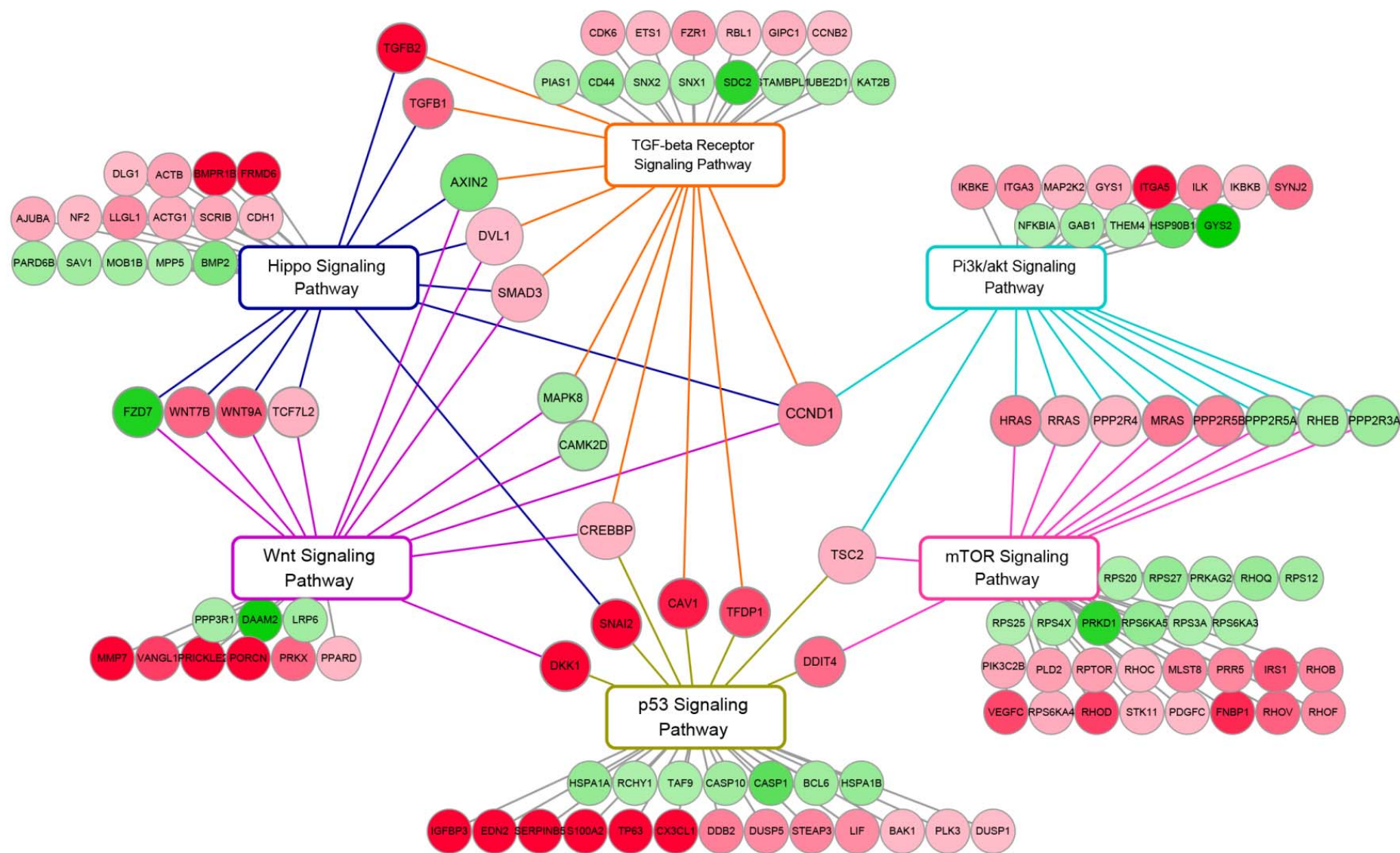
“Proximalization” of the peripheral lung in IPF

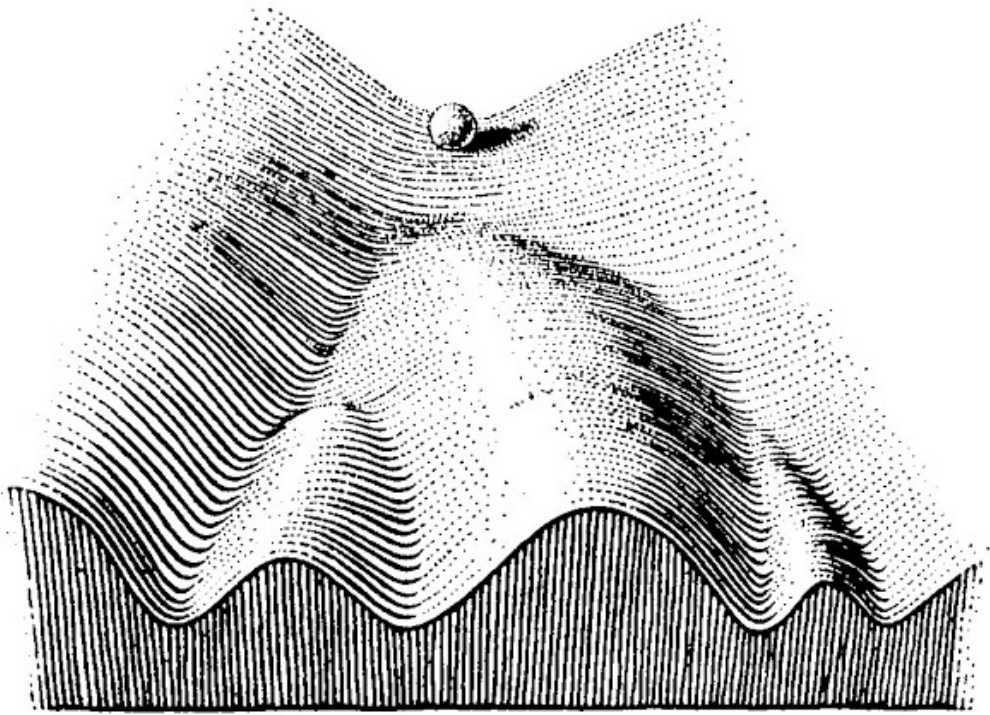


Activation of Cytokine and Growth Factor Signaling in IPF



Prediction of Abnormal Cell Signaling Driving IPF





The Epigenetic Landscape - Waddington 1957

Monogenic Diseases of Surfactant Homeostasis

ABCA3: Surfactant lipid transport in AT2 cells

SFTPB: Intra cellular packaging and function of surfactant

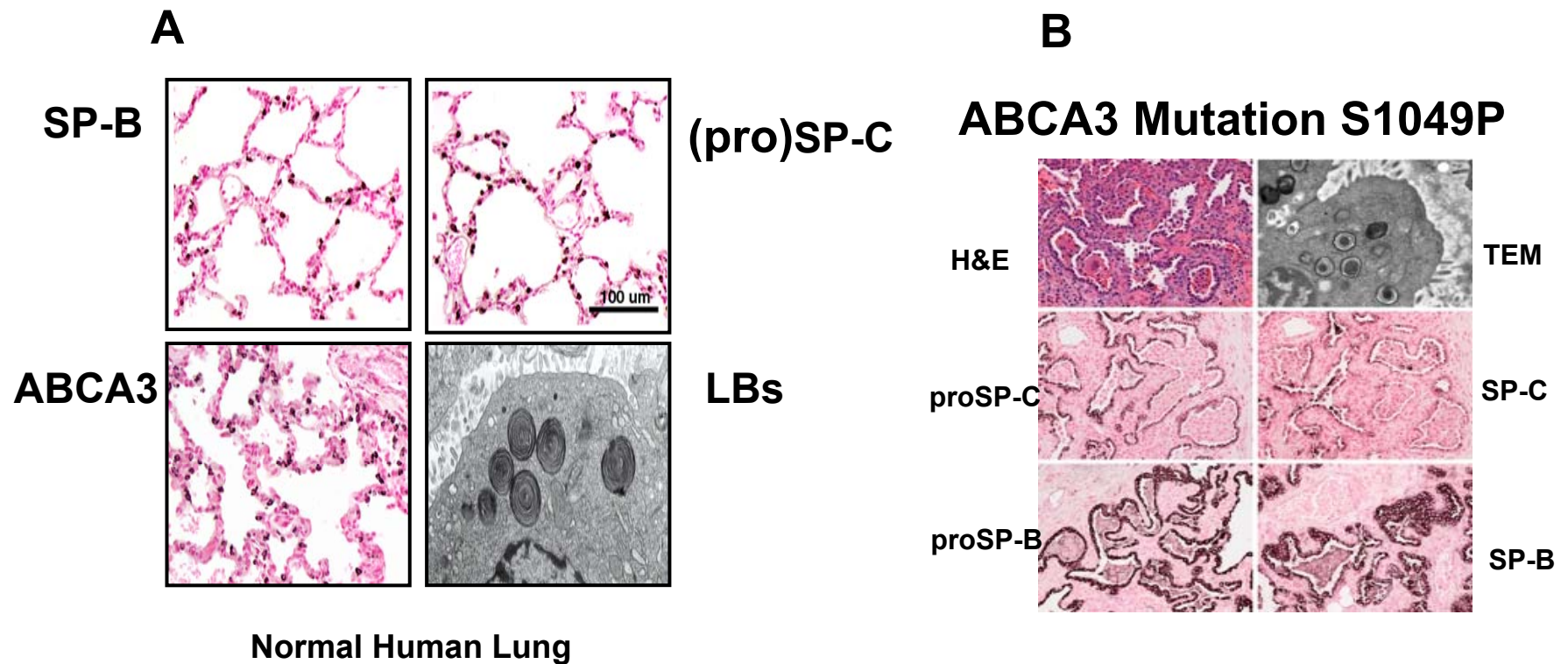
SFTPC: Surfactant function / host defense

SFTPA: Host defense, surfactant structure

NKX2-1: AT2 cell differentiation and function

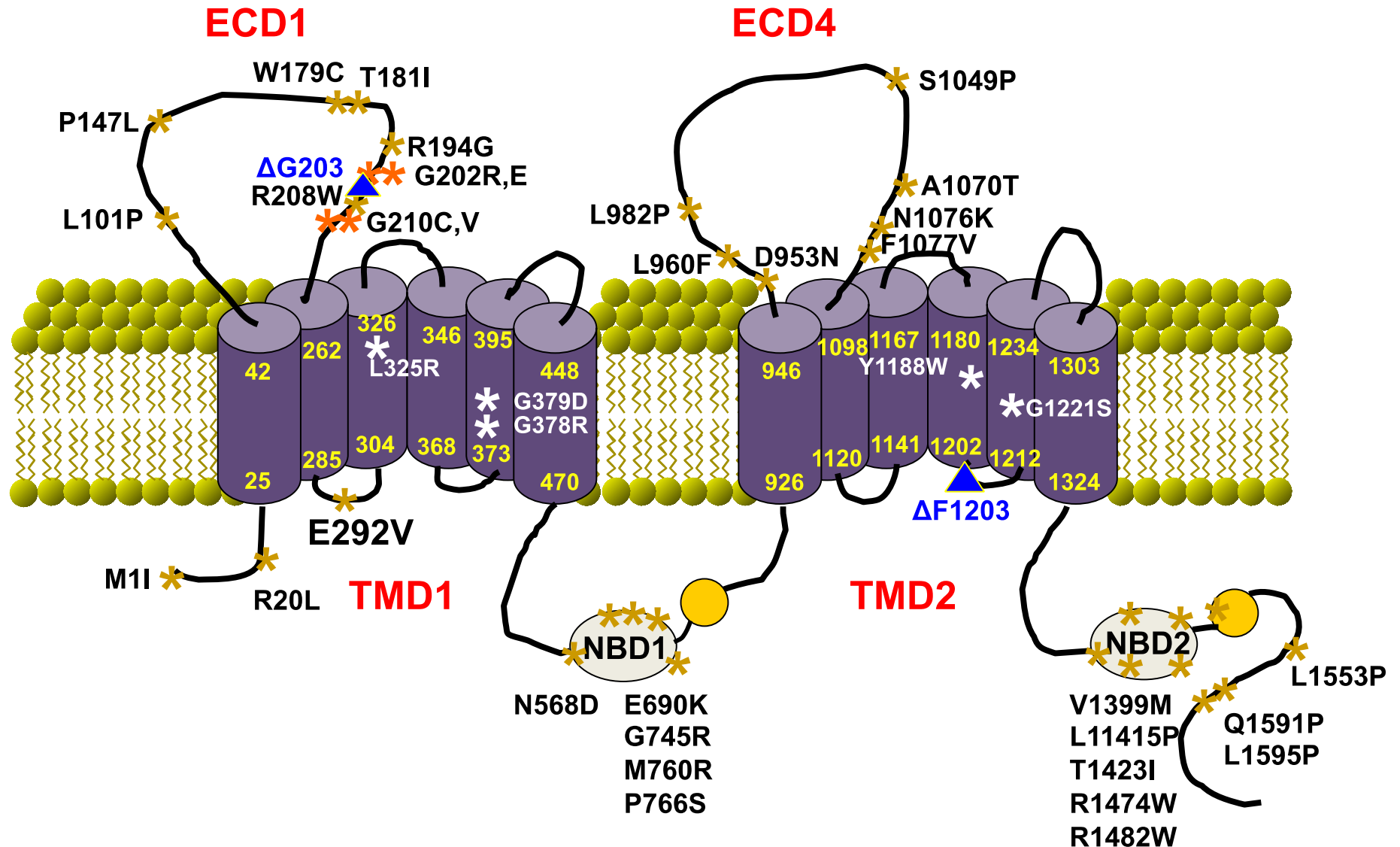
GMCSFa/b Receptors: Hereditary PAP/Macrophage

Histopathology of ABCA3 Deficiency



Partial Map of ABCA3 Mutations

Larry Noguee –Johns Hopkins

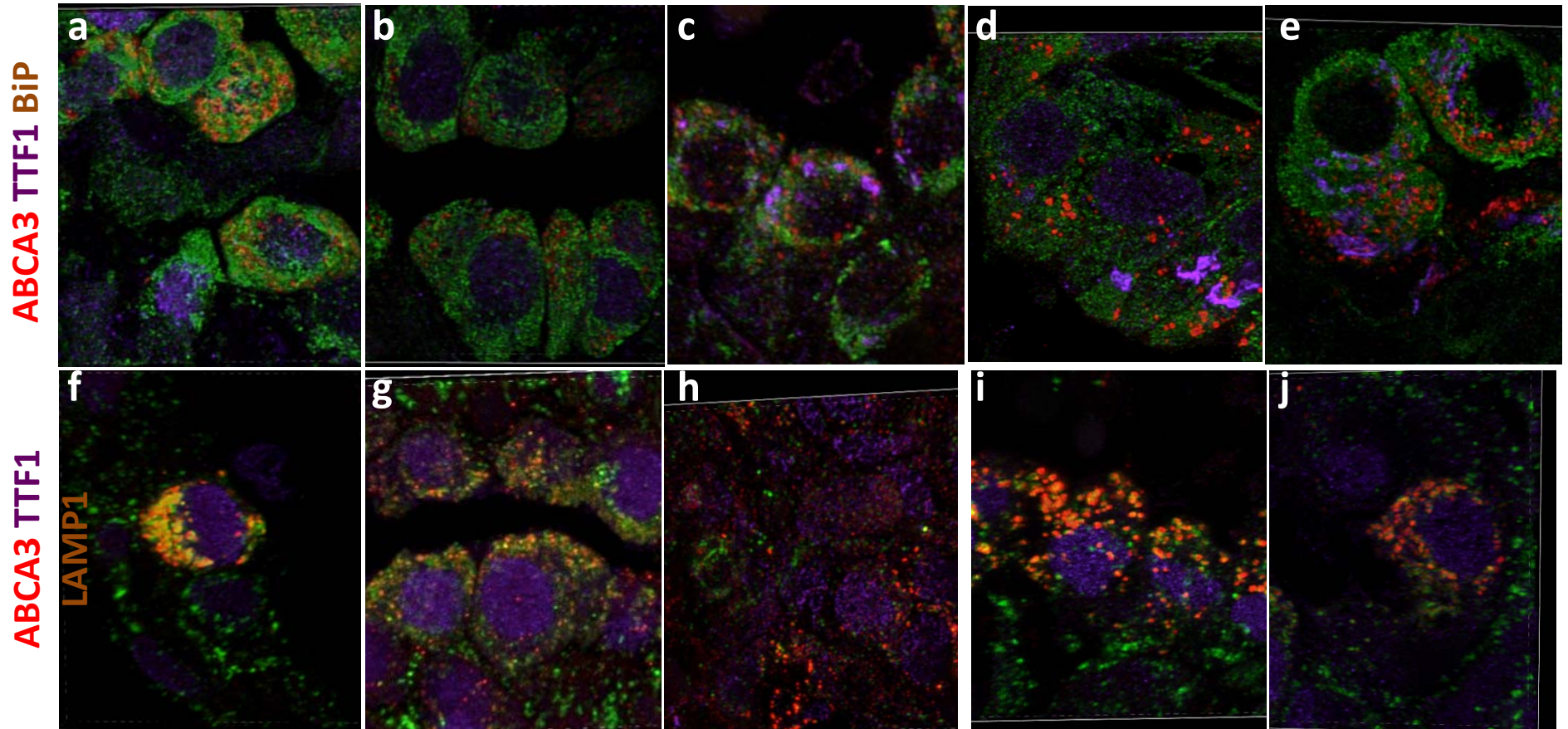


Current Classification of ABCA3 Mutations

Clinical Phenotype	Type I Mutations Impaired Processing / Trafficking	Type II Mutations Impaired ATP Hydrolysis / Lipid Transport
	Localized to ER	Localized to LB-like vesicles
NRF (11)	L101P (1,2,6) L982P (2) W1142X (2) L1553P (2) Gly1518Val*1 (2) Q215K (9,10)	R43L (6,8) R295C (severely impaired)(5) N568D (1,2,4) E690K (severely impaired) (4) T1114M (severely impaired) (4) G1221S (1, 2) L1580P (2)
chILD (11)	R280C (partial) (6) Q1591P (2)	E292V (moderately impaired) (4,9,10) D253H (7) T1173R (7)

NRF, Neonatal Respiratory Failure; chILD, Children's Interstitial Lung Disease

Subcellular Localization of ABCA3 Mutant Proteins



Expression of ABCA3 in human lungs. Confocal images were acquired by oversampling in x, y and z, deconvolved by landweber 3D. Control (**a and f**) and ABCA3 mutants S1049P/S1049 (**b and g**), Q1591P/Q1131R (**c and h**), W1142X/W1142X (**d and i**), and L982P/G1221S (**e and j**) stained with ABCA3 (red), TTF1 (purple) and the ER marker GRP78 (BiP) (green **top row panels a-e**) or lysosome marker LAMP1 (green **bottom row panels f-j**).

Progenitor Cell Translational Consortium -

Towards Correction of Monogenic Pulmonary Disorders

Collaborative Multi-PI Program

Jeff Whitsett – CCHMC

Ed Morrissey – University of Pennsylvania

Darrell Kotton – Boston University

F. Session Cole & Jennifer Wambach – Washington University

Larry Noguee – The Johns Hopkins University

Charles Ansong – Pacific Northwest National Laboratory

Towards Pathogenesis and Treatment of Monogenic Disorders of Surfactant Homeostasis

Model Human Surfactant Disorders: Mice, iPSC's, Organoids

Identify AT2 cell progenitors

Phenotype human/mouse AT2 s, iPSC's, Progenitors

Target and Deliver: RNA's, c DNA's, Gene editing tools to AT2 cell progenitors.

Pathogenesis of ABCA3 related disease:

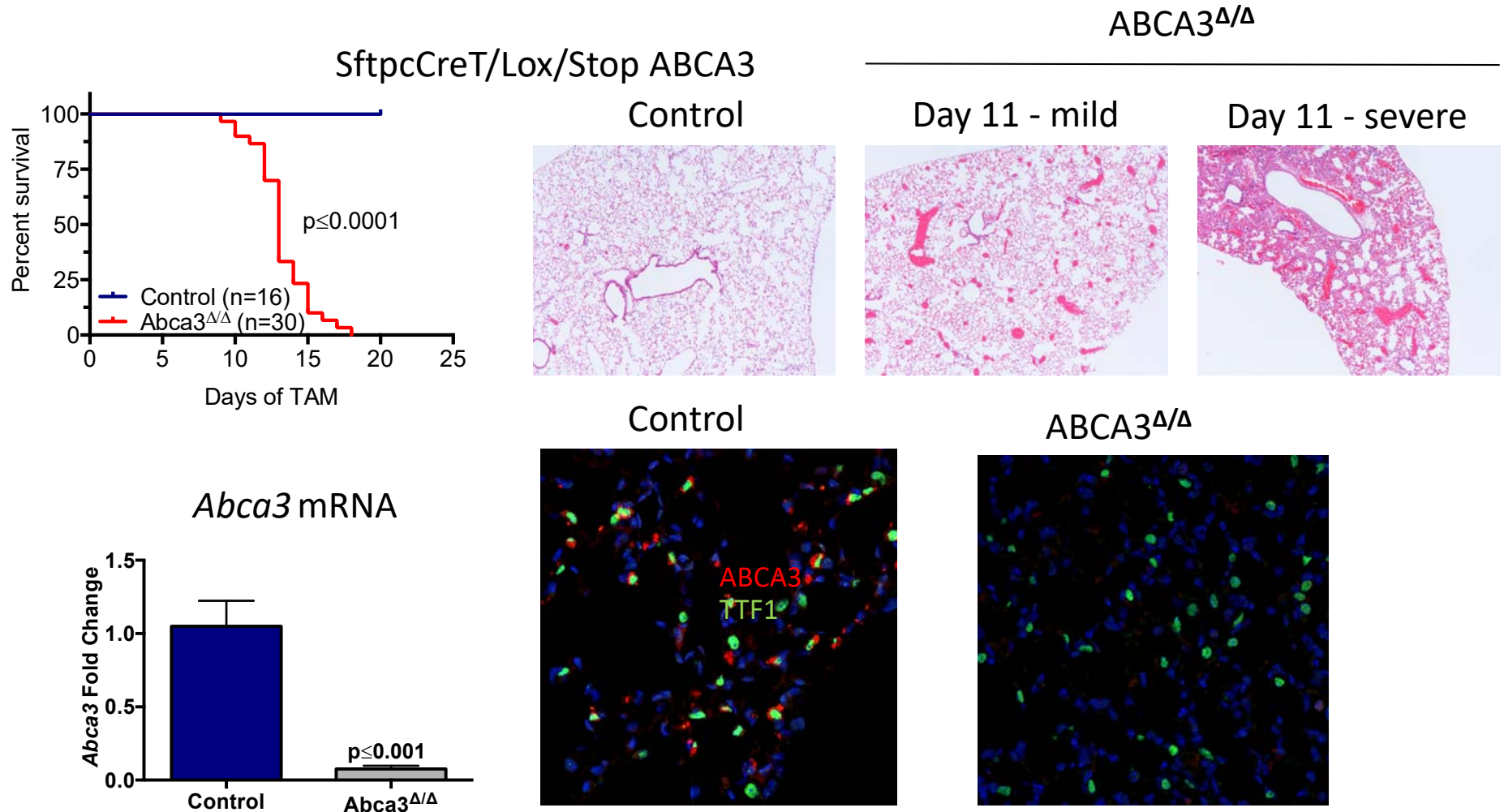
Molecular pathways: Modeling ABCA3 mutations

Proteomics, Lipidomics, Transcriptomics

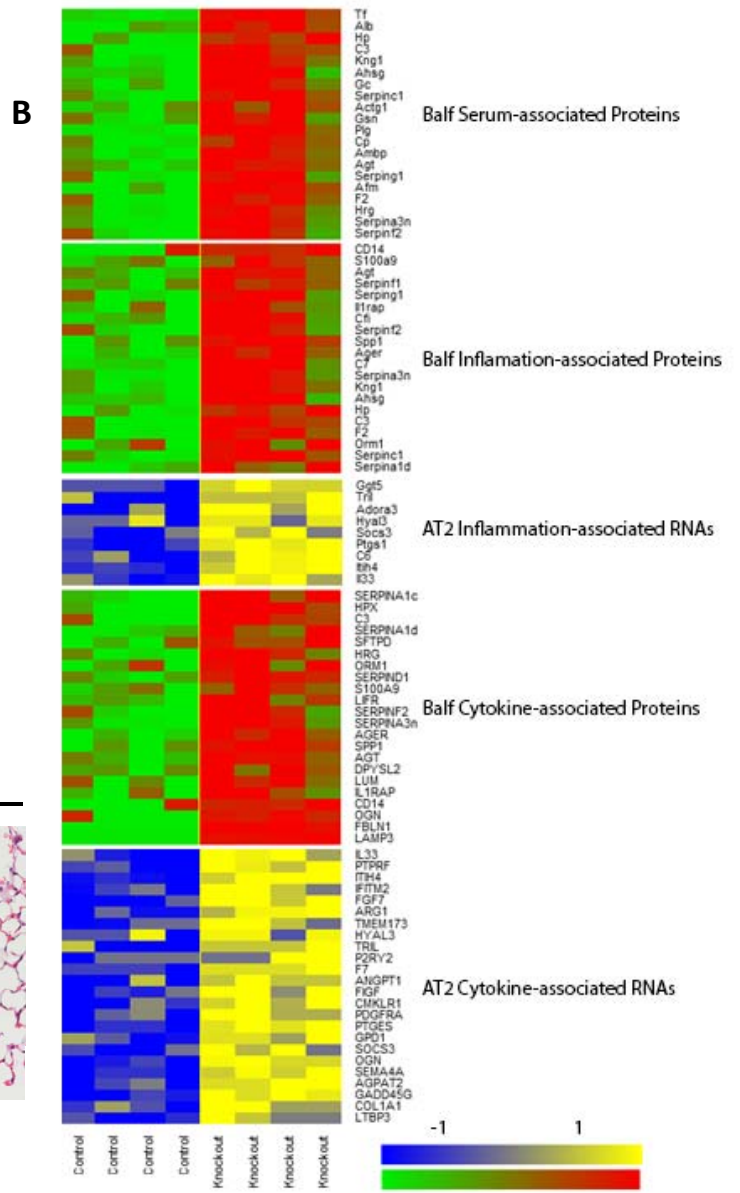
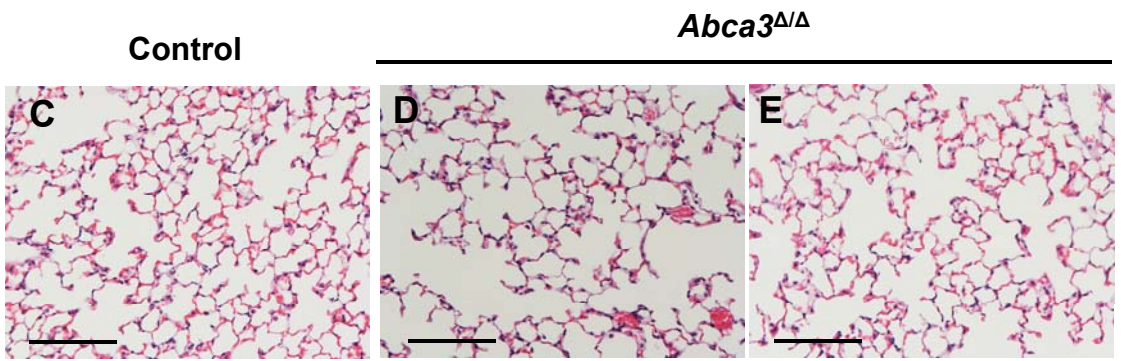
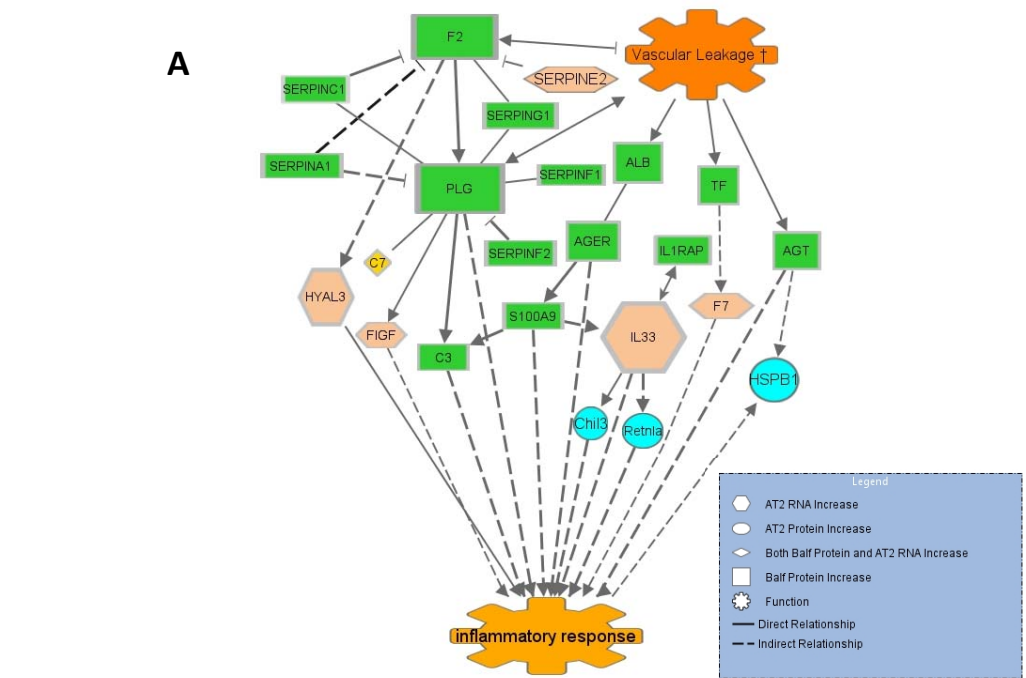
Mechanisms of tissue remodeling and lung dysfunction

Correction of ABCA3 disease: Replacement Editing

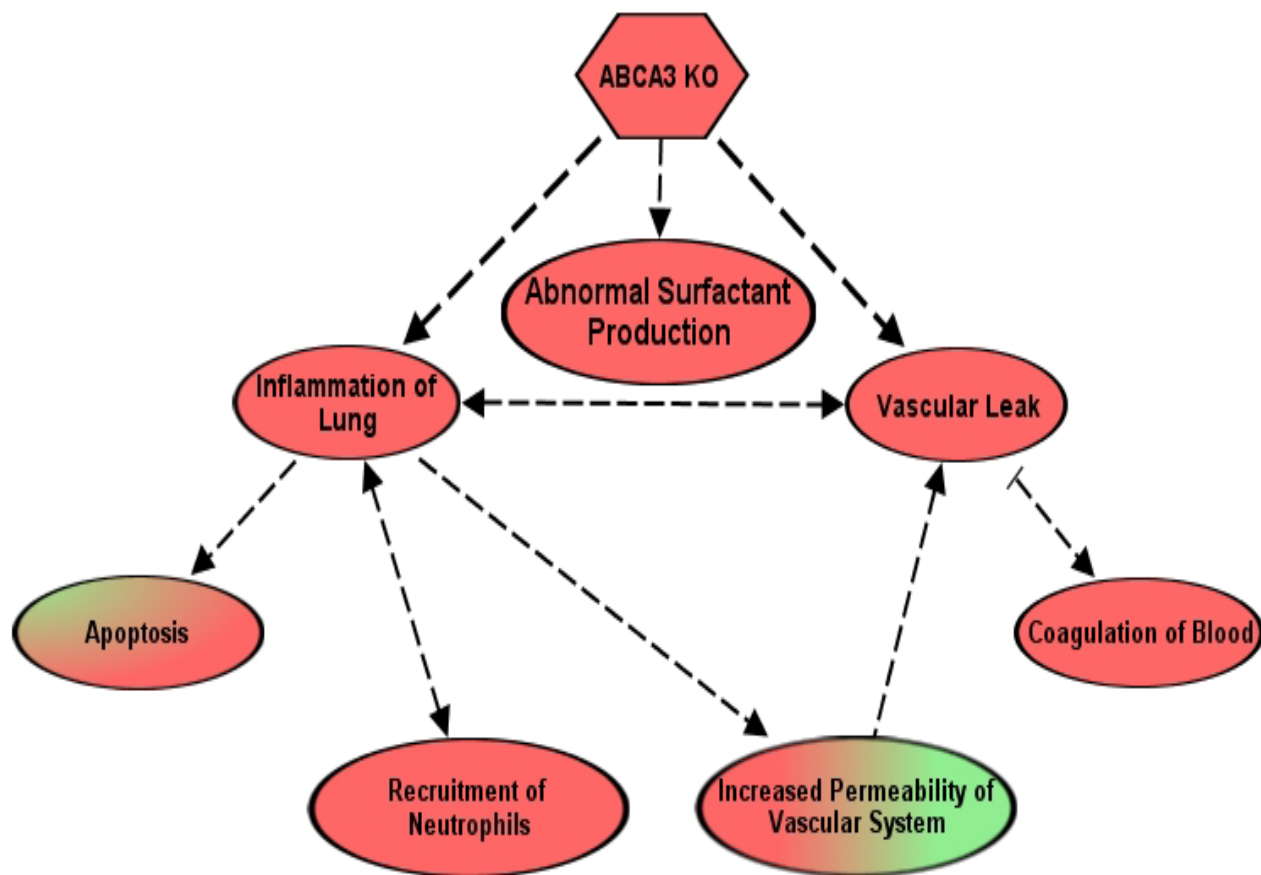
Conditional ABCA3 Deletion in Type II Cells Recapitulates Disease Phenotype in Mice



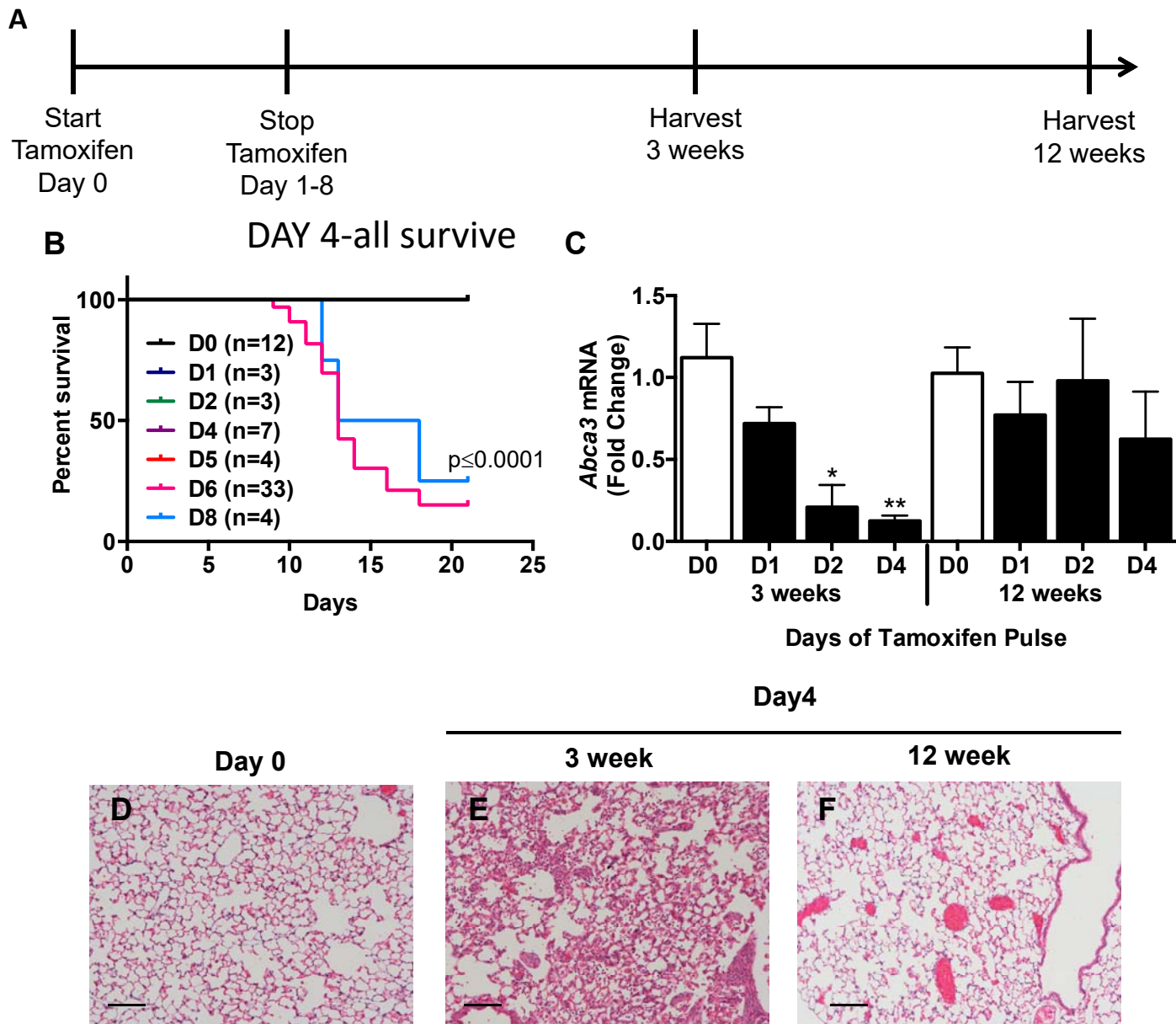
Deletion of ABCA3 initiates inflammation and capillary leak



Pathogenesis of Respiratory Failure in ABCA3 Deficiency

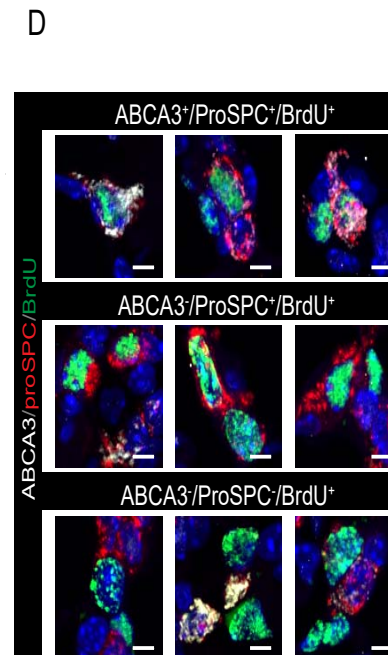
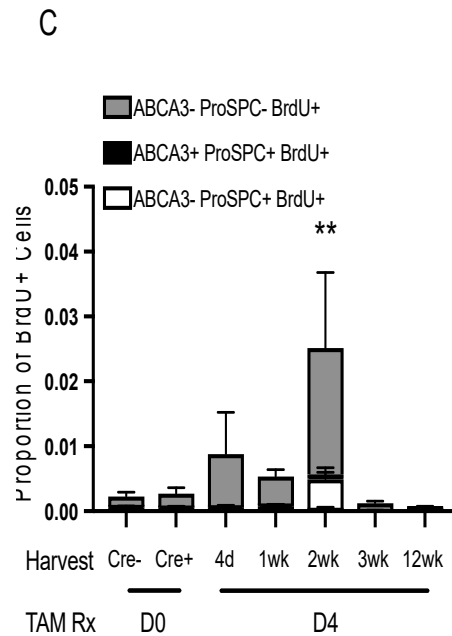
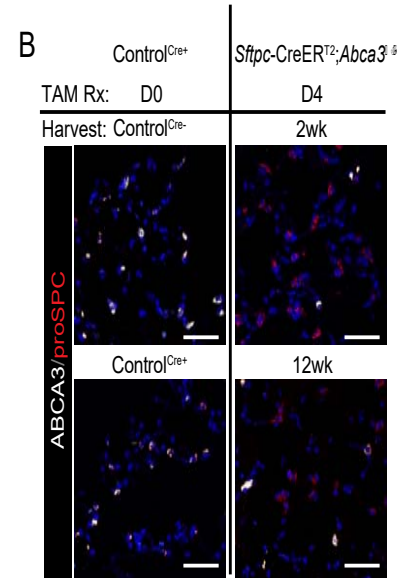
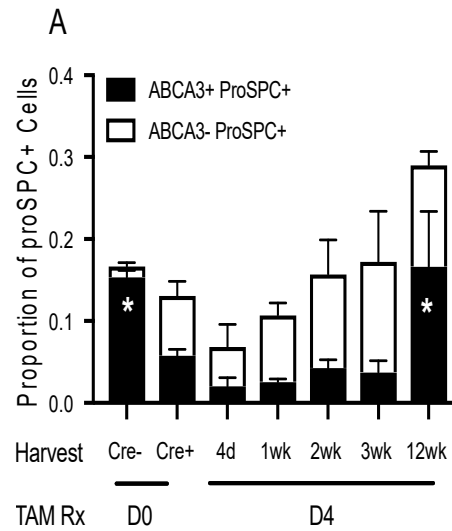


Remarkable Regeneration of the Peripheral Lung from ABCA3 Sufficient Cells

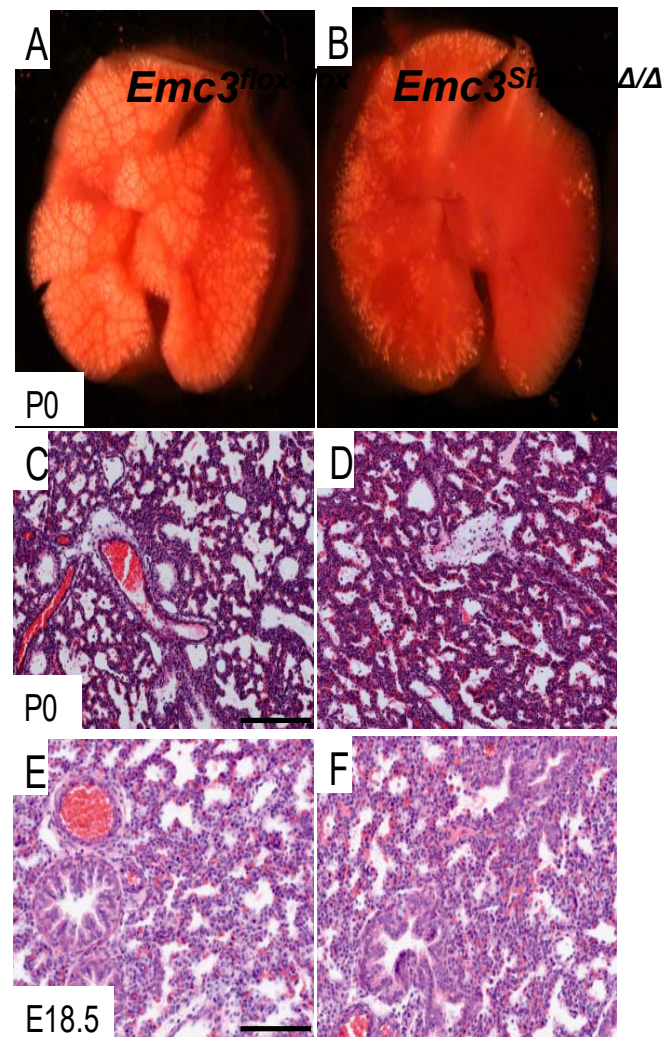


Complete

Alveolar Repair after Deletion of ABCA3

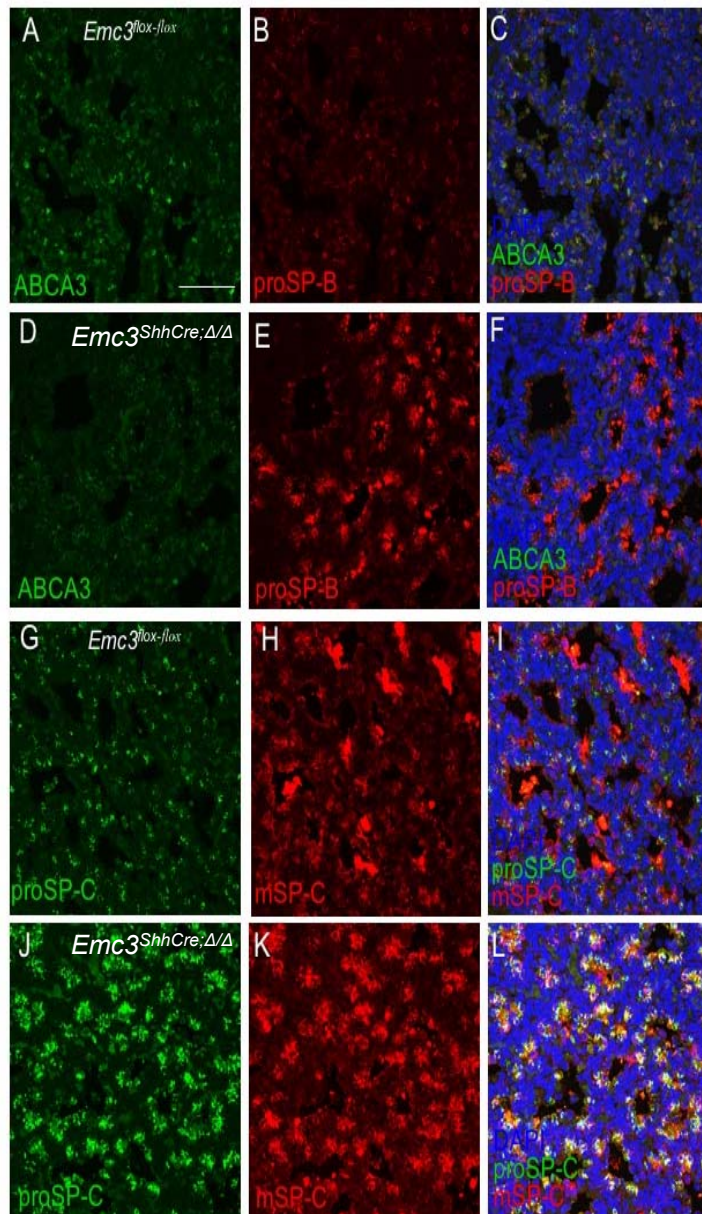


Cellular Mechanisms Controlling ABCA3/SP-B/SP-C Trafficking and Processing

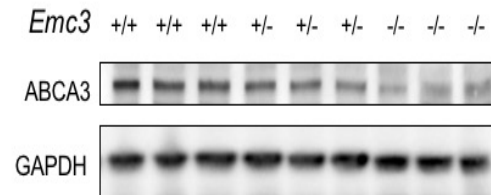


Respiratory Failure in EMC3 Deficient Mice

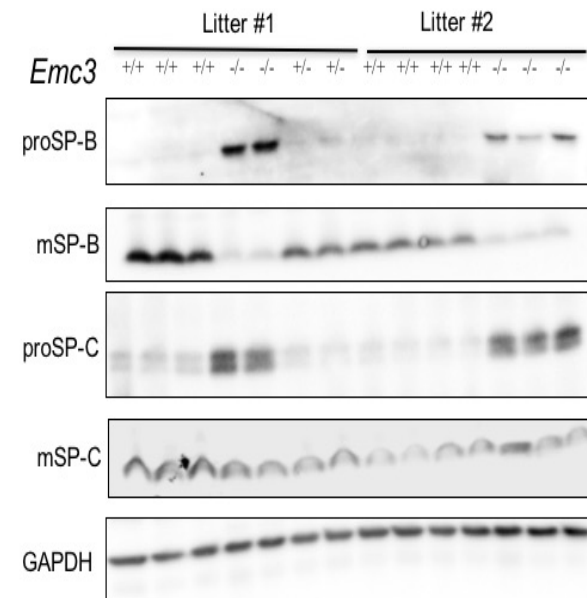
Decreased ABCA3 and Abnormal Surfactant Protein Processing in *Emc3*^{ShhCre;Δ/Δ} Mice



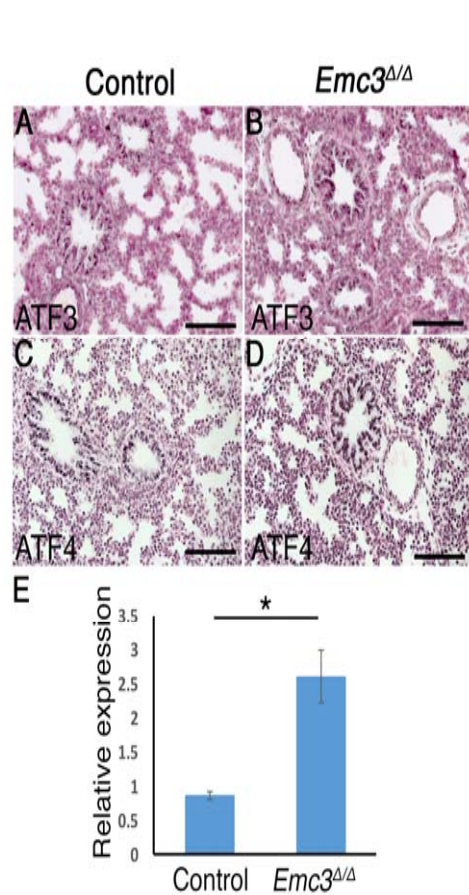
M



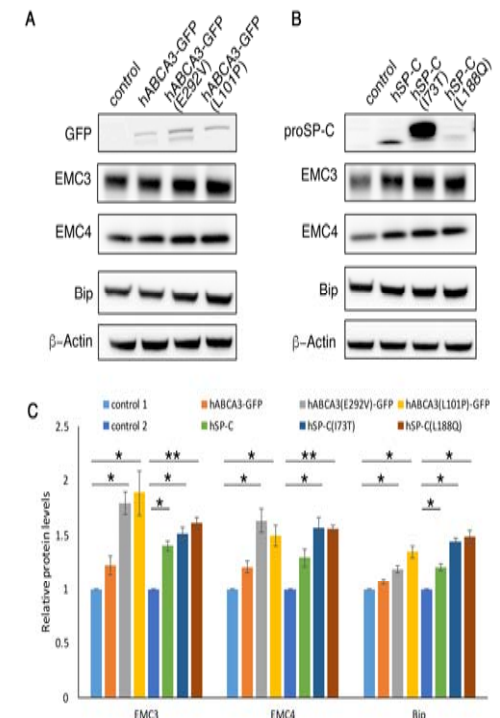
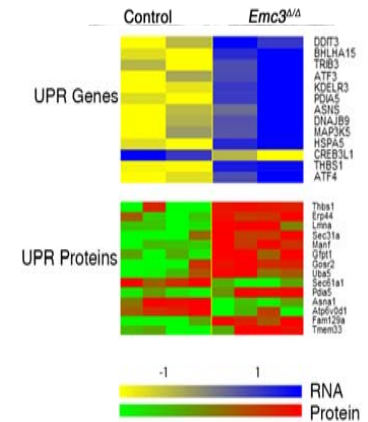
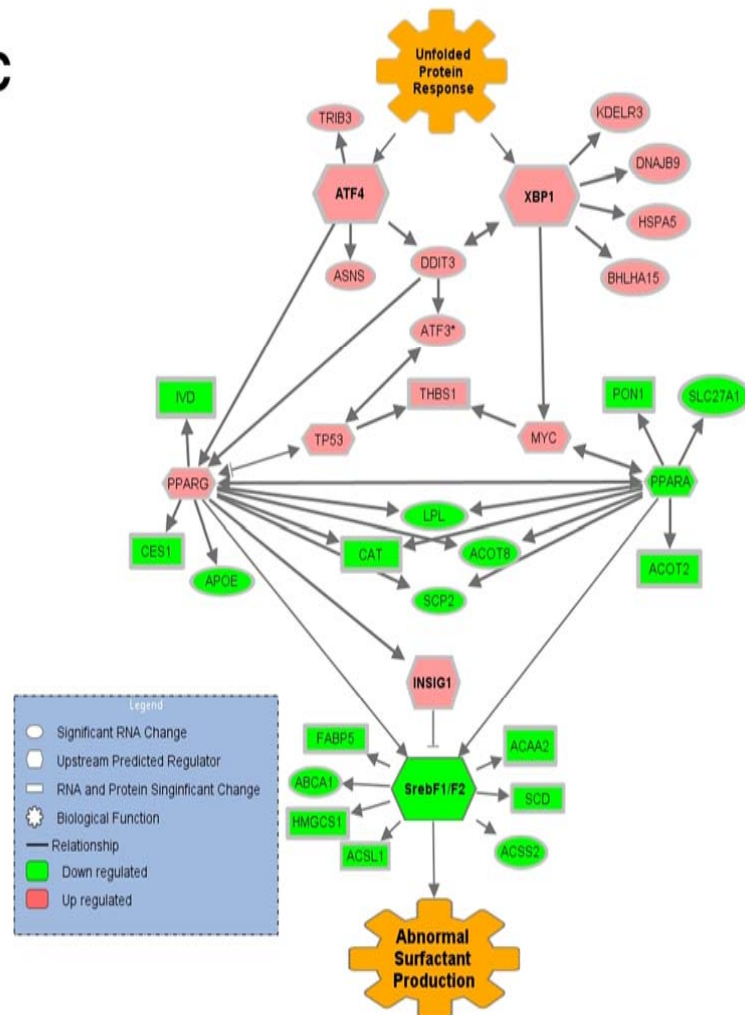
N



Role of EMC3 in UPR Modeled by Integration of RNA Sequencing and Proteomics



C



Genetic Basis of Disease

Model Systems

Patient Tissue
Organoids
IPSCs

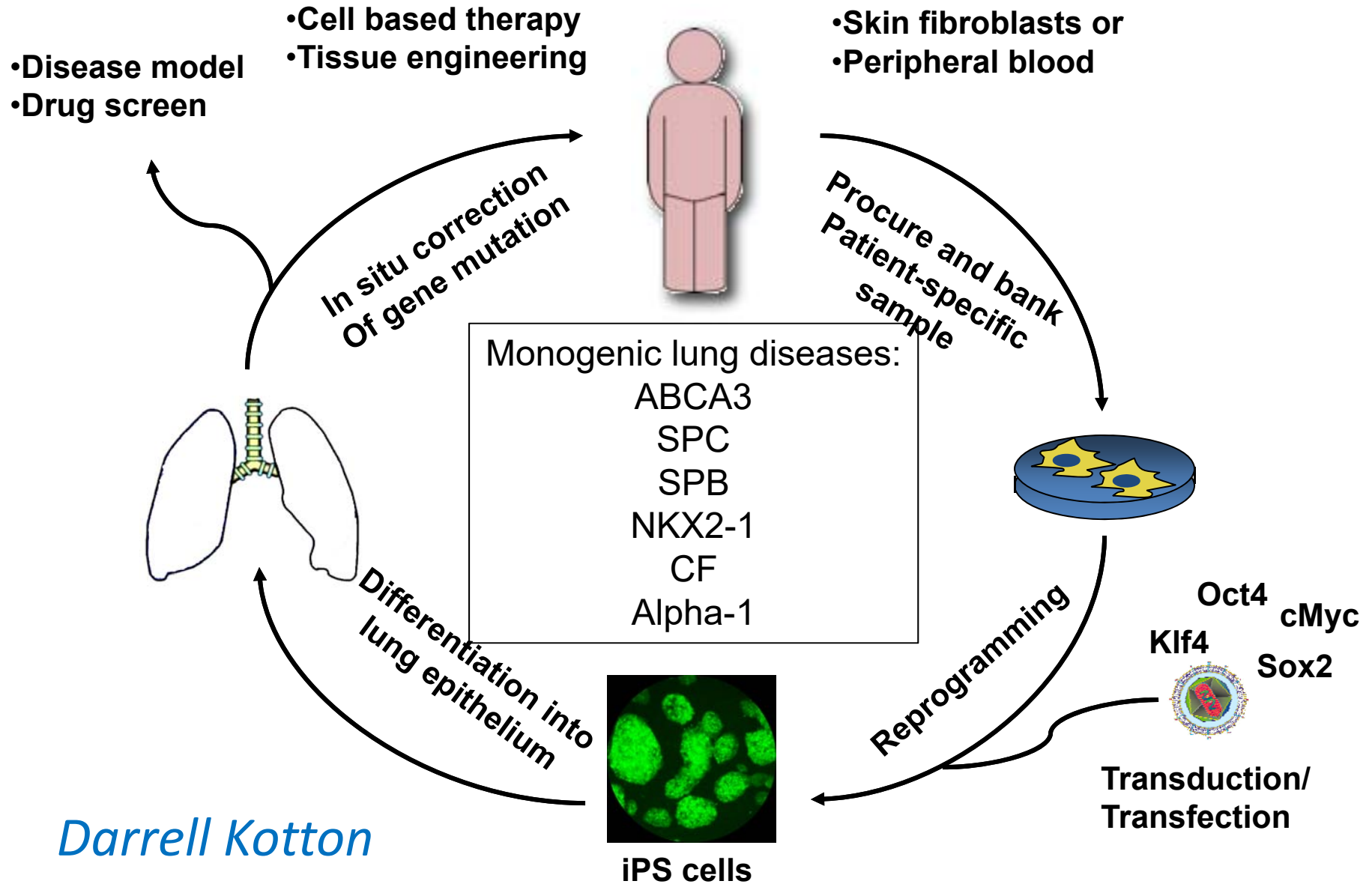
Advancing Technologies

“Systems Biology”

Mechanisms of Disease Pathogenesis

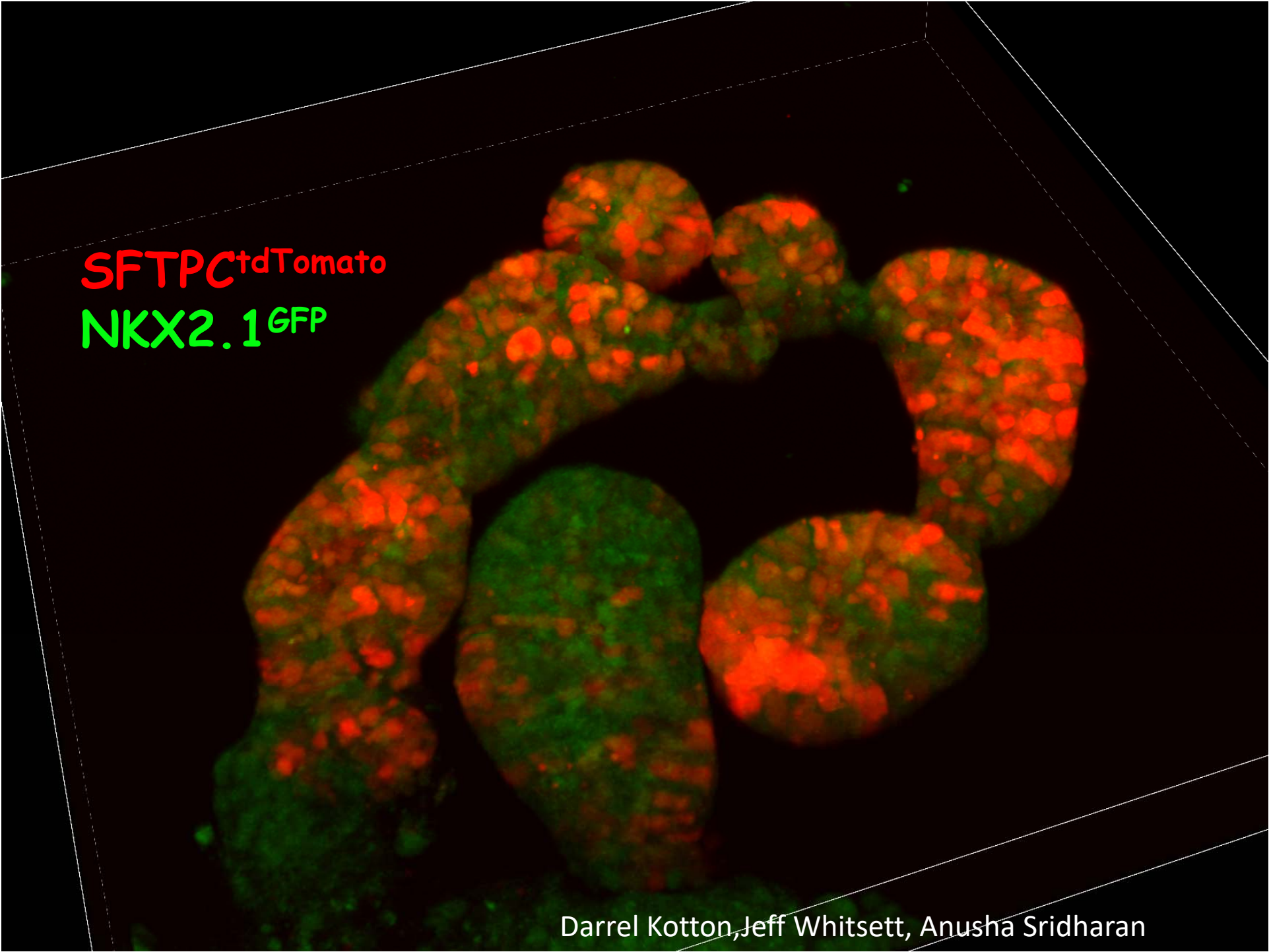
DX and RX

IPSCs to Model Disorders of Surfactant Homeostasis



Darrell Kotton

SFTPC^{tdTomato}
NKX2.1^{GFP}



Darrel Kotton, Jeff Whitsett, Anusha Sridharan

Vectors for Gene Transfer to the Respiratory Epithelium

AAV9/2 and AAV6/2: Single RNA's, Cas9, SP-C/SP-B

BOCA virus: ABCA3 ~ 5Kb

Lenti: ABCA3 and mutants

SNIMs: Stabilized, Modified RNA's: Cas9 sg RNA's

Can We Correct Monogenic Disorders of Surfactant Homeostasis?

Lung MAP and “Single Cell-NexGen Sequencing to Identify
Mechanisms Mediating Lung Formation Prior to Birth”

Pulmonary Biology



Yan Xu ,Yina Du

Anusha Sridharan

Joseph Kitzmiller

Minzhe Guo /XiaoFang Tang/Xinhua Lin

Anna-K Perl

Jim Bridges/Tara Rindler/John Snowball

Developmental Biology

Division of Bioinformatics

Bruce Aronow

Phillip Dexheimer

Steven Potter

Eric Brunskill

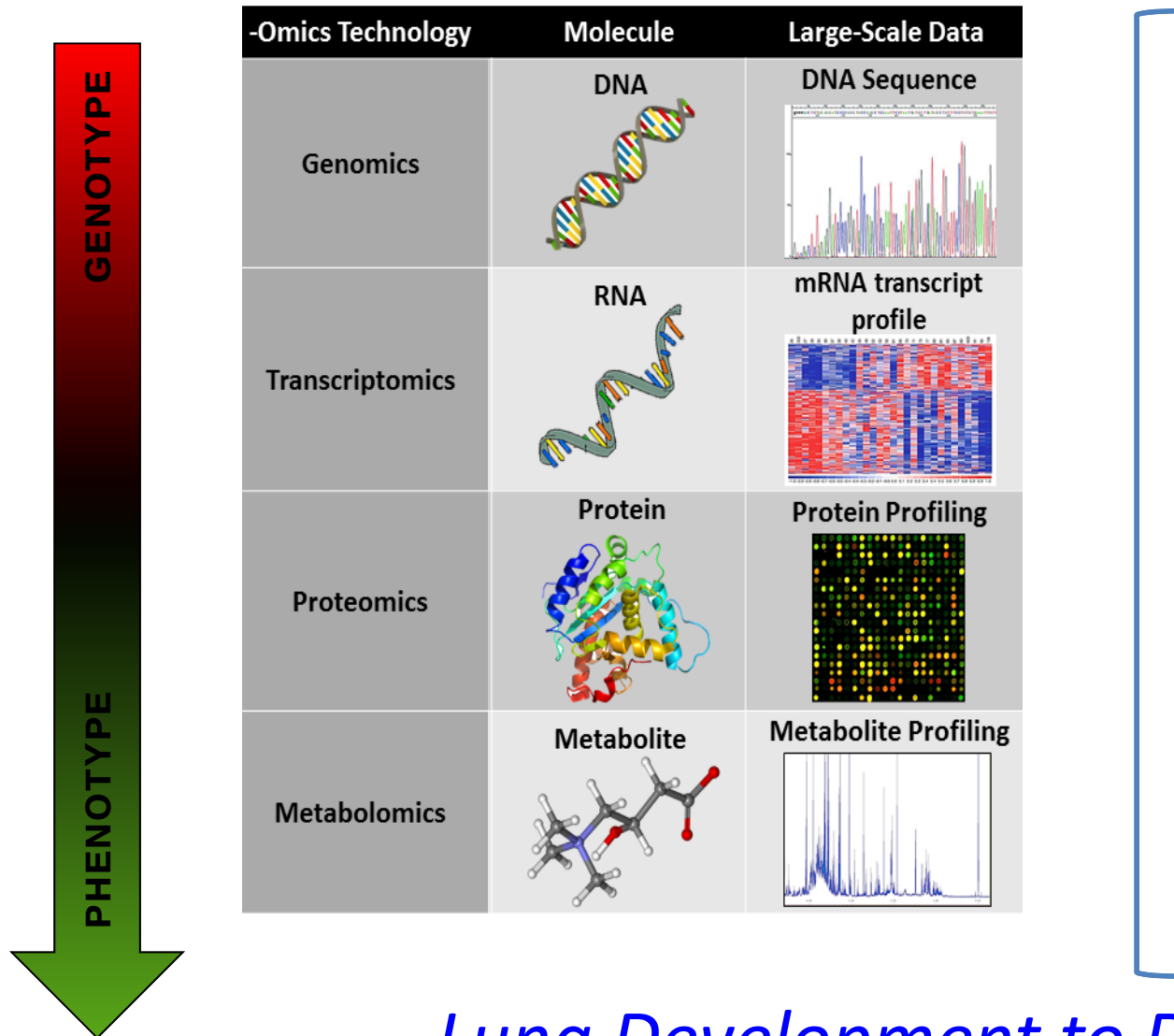
Matthew Kofron

DNA Core CCHMC

Imaging Core CCHMC

LungMap“omics”: Mouse and Human Lung

Data Integration Access and Display



Lung Development to Disease

Thanks!

LungMAP

Coordinating Center

Duke

Scott Palmer

Cliburn Chan

Laura Johnson

Jerry Kirchner

Carol Bova Hill



National Heart, Lung,
and Blood Institute

Research Centers

CCHMC

Jeff Whitsett

Steve Potter

Bruce Aronow

Yan Xu

PNNL

Richard Corley

Charles Ansong

UAB

Namasivayam

Ambalavanan

N Kaminski

J Hagood

USC

David Warburton

Wei Shi

Scott Fraser

Rex Moats

URMC – Tissue Procurement

Gloria Pryhuber

Ravi Misra

Progenitor Cell Translational Consortium:NHLBI